

# **Inflammatory activation after experimental cardiac tamponade and the effects of complement C5a inhibition**

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**Ph.D. Thesis**

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### List of full papers related to the subject of the thesis

1. **Vass A**, Süveges G, Érces D, Nógrády M, Varga G, Földesi I, Futakuchi M, Imai M, Okada N, Okada H, Boros M, Kaszaki J: Inflammatory activation after experimental cardiac tamponade. *Eur Surg Res* 51:1-13, 2013.DOI: 10.1159/000352089 IF 0.75
2. Érces D, Nógrády M, Nagy E, Varga G, **Vass A**, Süveges G, Imai M, Okada N, Okada H, Boros M, Kaszaki J: Complement c5a antagonist treatment improves the acute circulatory and inflammatory consequences of experimental cardiac tamponade. *Crit Care Med* 41: 2013. (accepted for publication) DOI: 10.1097/CCM.0b013e31828a6768 IF 6.123

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### Other publications

1. Nemes A, Forster T, Ungi I, Nagy V, **Vass A**, Pálinkás A, Varga A, Csanády M. The coronary flow velocity reserve measured by stress transoesophageal echocardiography evaluates the success of coronary interventions--results of a 5-year follow-up. *Scand Cardiovasc J.* 39: 286-292, 2005. IF:0.757
2. Rosztóczy A, **Vass A**, Izbéki F, Kurucsai G, Róka R, Horváth T, Lonovics J, Forster T, Wittmann T: Savas gastrooesophagealis reflux által provokált coronariaspasmus kórképe. *Magy Belorv Arch.* 61: 203-206, 2006. IF: 0
3. Rosztóczy A, **Vass A**, Izbéki F, Nemes A, Rudas L, Csanády M, Lonovics J, Forster T, Wittmann T: The evaluation of gastro-oesophageal reflux and oesophagocardiac reflex in patients with angina-like chest pain following cardiologic investigations. *Int J Cardiol.* 118: 62-68, 2007. IF: 1.765
4. Csanády M, Tóth F, Hogue M, **Vass A**, Sepp R, Csanády M Jr, Czigner J, Kiss JG, Jóri J, Forster T. Hearing disturbances in hypertrophic cardiomyopathy. Is the sensorineural disorder neurogenic or myogenic? *Int J Cardiol.* 116: 53-56, 2007. IF:1.765

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## **List of abbreviations**

|                 |  |
|-----------------|--|
| AcPepA          | acetyl-peptide-A (complement C5a antagonist) |
| big-ET          | big endothelin                               |
| CO              | cardiac output                               |
| CVP             | central venous pressure                      |
| ET-1            | endothelin-1                                 |
| EVLWI           | extravascular lung water index               |
| GEDV            | global end-diastolic volume                  |
| GEF             | global ejection fraction                     |
| HMGB1           | high-mobility group box protein-1            |
| HR              | heart rate                                   |
| IHC             | immunohistochemistry                         |
| LHEDV           | left heart end-diastolic volume              |
| LV              | left ventricular                             |
| MAP             | mean arterial pressure                       |
| MPO             | myeloperoxidase                              |
| NO              | nitric oxide                                 |
| NO <sub>x</sub> | nitrite/nitrate                              |
| PAP             | pulmonary arterial pressure                  |
| PiCCO           | pulse contour cardiac output                 |
| PMN             | polymorphonuclear neutrophil                 |
| PRSW            | preload recruitable stroke work              |
| PVRI            | pulmonary vascular resistance index          |
| RHEDV           | right heart end-diastolic volume             |
| RV              | right ventricular                            |
| SMA             | superior mesenteric artery                   |
| SV              | stroke volume                                |
| SVRI            | systemic vascular resistance index           |
| TNF- $\alpha$   | tumour necrosis factor-alpha                 |

## SUMMARY

Cardiogenic shock is common in patients with cardiac or extra-cardiac filling disorders. Cardiac tamponade is a life-threatening medical emergency situation where the pump failure is caused by effusions of blood, pus or fluids. Drainage of the pericardial sac through pericardiocentesis or surgical pericardiotomy is the first choice of treatment, but supportive medical therapy should be considered following or parallel to the relief of the intrapericardial pressure, with a view to avoiding low flow-induced peripheral complications. Moreover, it is recognized that splanchnic macro- and microcirculatory dysfunctions are linked to local and systemic inflammatory activation, and the increased afterload may subsequently worsen the hypoperfusion of the peripheral organs and result in an increased oxygen demand of the myocardium. The general purpose of the research reported in the dissertation was to characterize the major components of the pro-inflammatory profile of the post-tamponade phase, in association with the changes in overall haemodynamics, and to outline a possible therapeutic route to reverse the cardiac tamponade-caused detrimental consequences in a clinically relevant large animal model. We considered that a better understanding of the elements of these mechanisms may lead to new prospects for interventions designed to dampen or reverse the secondary detrimental consequences of cardiac failures.

In Study I, we hypothesized that acute failure of the myocardial pump function is accompanied by significant inflammatory activation, which can play important roles in further gastrointestinal complications. We investigated the immediate effects of cardiac tamponade on the systemic and peripheral circulations and characterized the major components of the pro-inflammatory profile of the post-tamponade phase, in association with the changes in overall haemodynamics. We identified characteristic macrohaemodynamic changes, together with apparent signs of a splanchnic inflammatory reaction after relief of the tamponade. The evidence further suggests that the activation of the complement and endothelin systems may be significant components of the inflammatory cascade that is activated in porcine pericardial tamponade. It is currently widely accepted that inflammatory activation plays a decisive role in low-flow conditions, although the potential of 'anti-inflammatory' compounds to prevent or cure hypoperfusion-induced *in vivo* processes is very limited. With this background, we hypothesized that early inhibition of complement C5a might well reduce the adverse cardiac and inflammatory consequences after relief of the cardiac tamponade. In Study II, during the acute phase of cardiac tamponade in our clinically relevant large animal model we used a synthetic, antisense, 17 amino acid peptide acetylated at the N-terminal alanine (acetyl-peptide-A - AcPepA). Previous studies have shown that this compound is capable of binding directly to C5a peptide. We hypothesized that, through this mechanism, AcPepA can influence the early cardiac and inflammatory changes, and the results furnished evidence that the administration of AcPepA significantly limits the extent of the cardiac dysfunction in this scenario.

## **1. INTRODUCTION**

### **1.1. Circulatory shock**

Maintenance of adequate perfusion of vital organs is critical for survival. Shock may be defined as a state in which profound and widespread reduction of tissue perfusion leads to reversible, and then, if prolonged, irreversible cellular injury. Since a reduction of tissue perfusion is a central issue, it is important to consider the factors that control this vital function. Organ perfusion is dependent on an appropriate perfusion pressure, which in turn is determined by two variables, the cardiac output (CO) and the systemic vascular resistance (SVR). The second critical determinant of arterial pressure, the CO, is the product of stroke volume (SV) and heart rate (HR).

The classification of shock is usually based on the aetiology. Cardiogenic shock, extracardiac obstructive shock, hypovolaemic shock and distributive shock are major categories (Parrillo 1991). The final pathway leads to cell death, but some characteristics of shock are the same regardless of the underlying cause or pathogenesis. One common denominator of different forms of shock is a low CO. Patients with shock develop a severe decrease in CO and hence a low perfusion of vital organs. The survival depends on the correct early diagnosis and therapy.

Two basic mechanisms can lead to a decrease of CO in shock: a failure of preload (i.e. a diminished venous return) in consequence of a decreased blood volume (in hypovolaemic shock) or a decrease in the efficiency of the pump function of the heart. The latter occurs in cardiogenic shock, in which the primary failure is myocardial rather than peripheral. This can arise after acute myocardial infarction or as a result of pericardial tamponade. In pericardial tamponade, compression of the atria and the right ventricle causes an acute rise in the pericardial pressure (by the accumulation of blood in the pericardial sac), which hinders the diastolic filling of the heart (Tyson *et al.* 1984), thereby leading to a decreased CO.

### **1.2. Determinants of cardiac pump function**

The myocardial performance is dependent on the *preload*, *afterload*, (*HR*) and *contractility*, and is influenced by neurohormonal and local endothelial cell-derived factors. The cardiac pump function or performance depends on many parameters, which act in parallel, and normally exist to some degree. Examples of performance parameters include the CO, left ventricular (LV) SV, the rate of fibre shortening, the stroke work, the myocardial compliance and the ejection fraction.

1. The *preload* is a result of heterometric autoregulation. When the cardiac muscle is stretched, it tends to develop greater contractile tension on excitation (this is Starling's law of the heart). Thus, the SV tends to increase in proportion to the preload (diastolic filling).
2. The *afterload* is determined by the total peripheral vascular resistance (TPVR). A sudden rise in afterload causes an immediate decrease in ejection fraction and a rise in end-systolic volume (ESV). The compensation can occur in two ways. An increased ESV coupled with a normal venous return leads to an elevated preload, and the SV is maintained by Starling's law. In addition, contractility may increase secondary to an increased coronary perfusion pressure, the Gregg effect (Anrep and Saalfeld 1933, Gregg 1963). Thus, small changes in afterload have a minimal steady-state effect on performance as measured via the SV or CO, despite a significant initial effect.
3. *HR* increases are usually associated with small increases in contractile force. However, an increased HR is associated with a proportionally shorter diastolic filling time and for this reason the atrial contraction becomes far more important at high HR. It has been established that the heart contractility depends on increasing HR in most mammals. The ventricular myocardium has an inherent ability to increase its strength of contraction, independently of neurohormonal control, in response to an increase in contraction. In humans, this myocardial property causes the contractile force to rise, as the contraction frequency is increased from 60 to about 180-200 beats  $\text{min}^{-1}$ , and then to decline with further increase in frequency (the force-frequency relation) .
4. *Contractility* is often defined as the intrinsic ability of a cardiac muscle fibre to contract at a given fibre length. Changes in the ability to produce force during contraction result from different degrees of binding between myosin and actin filaments. The degree of binding that occurs depends on the concentration of  $\text{Ca}^{2+}$  in the cell. The cytosolic  $\text{Ca}^{2+}$  level is the determinant of the involved myocardial fibre number in the contraction process and the maximum velocity of myocardial fibre shortening. An increased contractility, positive inotropic effect is reflected in a higher myocardial fibre shortening velocity with a higher cytosolic  $\text{Ca}^{2+}$  concentration in systole: more troponin is activated from higher levels of  $\text{Ca}^{2+}$  with more actin-myosin cross-bridges in unit time, and ultimately the myocardial fibre contraction is more extensive and faster. The performance improves at any given preload and afterload.

Neural influences mean that the sympathetic discharge to the ventricles increases their contractility. Likewise, circulating epinephrine increases contractility. Other inotropic agents, such as  $\text{Ca}^{2+}$  sensitizers, phosphodiesterase inhibitors, cardiac glycosides, catecholamines and



endothelial cell-derived factors, also modify the myocardial contractility and/or  $\text{Ca}^{2+}$  availability.

### **1.3. Analysis of cardiac function**

Examinations of cardiac contractility in clinical practice would be extremely beneficial, but at present direct measurement is not possible. In experimental practice, there are few invasive techniques available with which to evaluate or calculate this parameter. The end-systolic pressure-volume or pressure-diameter relationship is a fundamental description of systolic cardiac mechanics, with especial regard to the preload recruitable stroke work (PRSW) relationship. PRSW is another index of contractility, which is perhaps less influenced by other parameters. Each pressure-volume/diameter loop is derived by caval vein occlusion. The slope of the derived linear relationship is a measure of contractility independent of the preload and afterload (Molnár *et al.* 2011).

In clinical practice, non-invasive measurements of LV dimensions, volumes, ejection fraction and other indices of the systolic and diastolic function can be carried out by ultrasound techniques. Transthoracic echocardiography has an important and established diagnostic role and has been used successfully to monitor segmental wall motion, ejection fraction, valvular function, and to measure cardiac cavities. The evaluation of the ejection fraction is important for risk stratification before any surgical intervention. However, the ejection fraction is influenced by preload and afterload alterations without any change in contractility. Depending on the loading conditions, hearts with a lower ejection fraction can produce a greater CO. Although roughly indicative of the cardiac reserve, the ejection fraction is an inconsistent marker of the overall cardiac function preoperatively.

Assessment of the LV function includes linear measurements, such as the LV internal dimension in diastole and systole, from which parameters such as fractional shortening could be derived. Two-dimensional echocardiography allows area measurements and derived volume calculations (i.e. Simpson rule measurements). Once the systolic and diastolic volumes have been determined, the SV can be calculated. The forward CO then equals the product of HR and SV. Assuming the absence of a mitral or aortic insufficiency, this can represent the CO. The application of Doppler echocardiography provides information on systolic flow parameters, and more recently the diastolic function. Doppler techniques have several advantages: they are non-invasive and simple, and provide continuous data. In contrast, they also have some important limitations, such as the relative inaccuracy (high inter/intraobserver variability) of the cross-sectional measurements in the LV outflow tract.

With the help of some sophisticated new methods, such as strain rate imaging, the ventricular function can be determined exactly, but from a practical point of view it is rather complicated. Another new method is automated endocardial border definition, used for automatic quantification of the instantaneous LV volume. The SV and ejection fraction can be calculated from the maximum and minimum volumes. A further refinement of this method in combination with the instantaneous determination of systolic pressure is that it can provide load-independent information regarding ventricular contractility through the creation of a pressure-volume loop (Gorcsan *et al.* 1994), although its routine use during the operation and subsequently in the intensive care unit is limited.

There are two prevalent *invasive* techniques in clinical practice, for the measurement and calculation of the preload and CO parameters by a thermodilution method. One of these techniques, which applies the pulmonary artery (Swan-Ganz) catheter, has been used for over 20 years. Pulmonary artery thermodilution and the subsequent clinically established transcatheter pulmonary thermodilution technique allow the measurement not only of the CO, but also of several other parameters reflecting the right (RV) and the biventricular function. Pulmonary artery thermodilution permits the measurement and calculation of the RV ejection fraction, the RV stroke volume, and the RV end-diastolic volume (Trepte *et al.* 2011). With the development of additional facilities, and particularly the continuous measurement of CO, the pulmonary artery catheter remains the mainstay in the bedside monitoring of myocardial performance in critically ill patients.

Another invasive technique is the Pulse Contour Cardiac Output (PiCCO) system, which uses volumetric, transcatheter pulmonary thermodilution. This technique is methodologically comparable and has been described as being a reliable and less invasive method for the measurement of CO (Reuter *et al.* 2010). It further enables an assessment of the global ejection fraction (GEF), describing the ratio of the SV to the global end-diastolic volume (GEDV). For the calculation of GEF, the SV is multiplied by 4 on the assumption that the GEDV represents the volume of all four heart chambers ( $GEF = 4 \times SV / GEDV$ ). The main advantages of the PiCCO system are the online function and the possibility of pulse contour analysis. A combination of the pulse contour CO and volumetric ejection fraction monitoring system permits calculation of the right heart and left heart end-diastolic volumes (RHEDV and LHEDV) (Végh *et al.* 2009).

Less invasive methods for continuous and rapid perioperative monitoring are clearly needed to detect changes in central haemodynamics in order to optimize the haemodynamic performance and oxygen delivery. A promising method is an ultrasound-based technique for

the measurement of blood flow velocity and aortic diameter simultaneously in the descending aorta, with the use of transoesophageal echocardiography with a probe containing two ultrasound transducers (Odenstedt *et al.* 2001).

#### **1.4. Cardiac tamponade and its circulatory effects**

Cardiogenic shock may occur in patients with cardiac or extracardiac filling disorders such as cardiac tamponade (Topalian *et al.* 2008, Bodson *et al.* 2011). Cardiac tamponade is a typical cause of obstructive shock. It is defined by an acute circulatory failure secondary to compression of the heart chambers by a pericardial effusion. Tamponade is a life-threatening condition that is diagnosed clinically by an elevated jugular venous pressure, hypotension and *pulsus paradoxus* in the setting of a pericardial effusion. The diagnosis is usually easy if shock is present, but it may be difficult if the effusion develops without obvious circulatory failure. Knowledge of the pathophysiology and the echocardiographic characteristics of this disease is therefore fundamental.

The incidence of cardiac tamponade is poorly documented. Numerous epidemiological studies have examined patients suffering from a pericardial effusion without focusing on tamponade. Interestingly, it was observed in a retrospective study of more than 10000 ambulatory patients without previous pericardial disease that the existence of a small pericardial effusion on transthoracic echocardiography was associated with an increased 1-year mortality rate from 11% to 26% (Mitiku and Heidenreich, 2011).

Studies establishing the cause of pericardial effusion have yielded mixed results, probably due to the different diagnostic techniques applied. In a single centre study, 106 patients were hospitalized for a large pericardial effusion, defined by a thickness greater than or equal to 20 mm in diastole, with or without a syndrome of cardiac tamponade; the causes found were cancer (36%), idiopathic (30%), infection (21%), myxoedema (8%), autoimmune diseases and vasculitis (5%). The reported incidence of acute pericardial tamponade is approximately 2% of penetrating trauma. In emergency situations, a haemopericardium is frequently encountered in patients with ascending aorta dissection or ventricular rupture following infarction. After cardiac surgery, the development of a pericardial haematoma behind the right atrium is a common cause of shock. However, its diagnosis may be difficult and requires transoesophageal echocardiography (Bodson *et al.* 2011). The increasing number of cardiac surgical interventions has resulted in a subsequent elevation in the overall number of iatrogenic pericardial tamponades. Further potential causes of cardiac perforation include

central line placement, pacemaker insertion, cardiac catheterization and electrophysiological ablation procedures (Valley and Fly, 2007).

A raised intrapericardial pressure can occur by three main mechanisms: increased fluid within the intrapericardial space (pericardial tamponade), an increased volume of the cardiac chambers (cardiac diseases), or an increased stiffness of the pericardium (constrictive pericarditis). In the normal situation, the pericardium has a small capacitance reserve that will accommodate only a small increase in cardiac chamber size and/or pericardial fluid volume (approximately 150-250 mL) before a significant increase in the pericardial pressure (Goldstein 2004).

The acute rise in pericardial pressure compresses the atria and the right ventricle and hinders the diastolic filling of the heart (Tyson *et al.* 1984), thereby leading to a decreased CO. Vasoconstriction is a general compensatory reaction in forms of shock involving low CO. It occurs as an appropriate response that serves to restore the declining arterial flow and pressure towards former values. The vasoconstriction is non-uniform, and the redistributed blood flow can maintain the blood supply for the heart and brain at a relatively normal level in the short run. The sympathetic vasoconstriction diverts the blood flow from organs with constricted arterioles to organs whose vessels constrict little under intense vasoconstriction. The resistance of the coronary circulation practically does not change, whereas the resistance of the skin, skeletal muscle and gastrointestinal circulations increases enormously (Bond and Green, 1983). This process is mediated through the activation of sympathetic nervous system and various humoral mediators (Kaszaki *et al.* 1989), leading to systemic vasoconstriction, tachycardia and fluid retention (Topalian *et al.* 2008). However, the price for the potentially beneficial effects of selective vasoconstriction is severe hypoxia in the underperfused organs (Bulkley *et al.* 1983, Bailey *et al.* 1987, Buerke *et al.* 2011). Moreover, the increased afterload may subsequently worsen the peripheral hypoperfusion and increase the oxygen demand of the myocardium.

Circulatory disturbances of cardiac origin fundamentally influence the perfusion of other organs, and especially the gastrointestinal tract. Gastrointestinal perfusion is often compromised early relative to other vascular beds in critical illness, major surgery or exercise due to the neurohumoral elements of circulatory redistribution. This complex response is partly mediated through the sympathetic nervous system (Chien 1967), but various humoral mediators also participate in the maintenance of vasoconstriction (catecholamines, the renin-angiotensin system and endothelins).

In cases of cardiac tamponade, drainage of the pericardial sac through pericardiocentesis or surgical pericardiotomy is the first choice for therapy (Seferovic *et al.* 2006), but removal of the pericardial fluid can lead to a reperfusion phenomenon with further peripheral reactions. As a consequence of hypoxia and reoxygenation, reactive oxygen radicals are generated and polymorphonuclear neutrophil (PMN) leukocytes are activated. The formation of these radicals may lead to disintegration of the cell membranes, structural damage and decreased cellular functions (Boros *et al.* 1989, Prasad *et al.* 1990). High-mobility group box protein-1 (HMGB1), released passively by necrotic and damaged cells, was recently identified as an important signal for leukocyte recruitment (Scaffidi *et al.* 2002). Further factors identified in the background of PMN leukocyte accumulation are an increased level of endothelin-1 (ET-1) formation and a decreased level of nitric oxide (NO) formation, which coexist in ischaemia-reperfusion syndromes (Boros 2003). ET-1, one of the most powerful endogenous vasoactive mediators, may contribute to the impairment of the microcirculation through its vasoconstrictor and pro-adhesive effects (Boros *et al.* 1998). Moreover, it results in histamine release from resident mast cells (Kaszaki *et al.* 2008), and influences the activation of the complement cascade (Soop *et al.* 2004).

### **1.5. A possible therapeutic route: the complement system**

Complement was first discovered in the 1890s when it was found to aid or “complement” the killing of bacteria by heat-stable antibodies present in normal serum. The complement system consists of more than 30 proteins that are present either as soluble proteins in the blood or as membrane-associated proteins. This system is composed of a network of proteins that play an important role in innate and adaptive immunity, ranging from the opsonization of pathogens, through chemoattraction to the removal of apoptotic and necrotic cells. Activation of the system is exquisitely regulated, and inappropriate activation due either to deficiencies in key complement proteins or to dysregulated activation has adverse consequences (Sarma and Ward 2011). Complement activation leads to a sequential cascade of enzymatic reactions (known as complement activation pathways), resulting in the formation of the potent anaphylatoxins C3a and C5a, which elicit a plethora of physiological responses that range from chemoattraction to apoptosis. These proteins are capable of cell lysis and the principal site of synthesis is the liver. Complement activation is known to occur through three different pathways: alternative, classical and lectin (Figure 1), involving proteins that mostly exist as inactive zymogens, which are sequentially cleaved and activated. Activation of the classical complement pathway occurs via C1, C4 and C2, and activation of

an alternative complement pathway via factor D, C3 and factor B. Both pathways lead to the activation of C3. The protein fragment C3b, split form C3, is necessary for activation of the terminal complement components, C5-9. These form the membrane attack complex. When inserted into cell membranes, they cause osmotic lysis of the cell. C3a and C5a bind to receptors on mast cells and basophils, resulting in the release of histamine and other mediators of anaphylaxis.

C5a is a potent chemoattractant for PMNs and monocyte-macrophages. C5a is a 74 aminoacid peptide generated from C5 during complement activation. C5a acts efficiently as an anaphylatoxin, stimulating cells such as leukocytes and endothelial cells, and is also a potent chemotactic factor for PMNs and other inflammatory cells bearing the C5a receptor. C5a is therefore considered to be one of the most potent inflammatory mediators. Inflammatory cells respond to nanomolar concentrations of C5a with intracellular  $\text{Ca}^{2+}$  mobilization, stimulation of chemotaxis, aggregation, degranulation and the production of superoxide anions (Fujita *et al.* 2004).

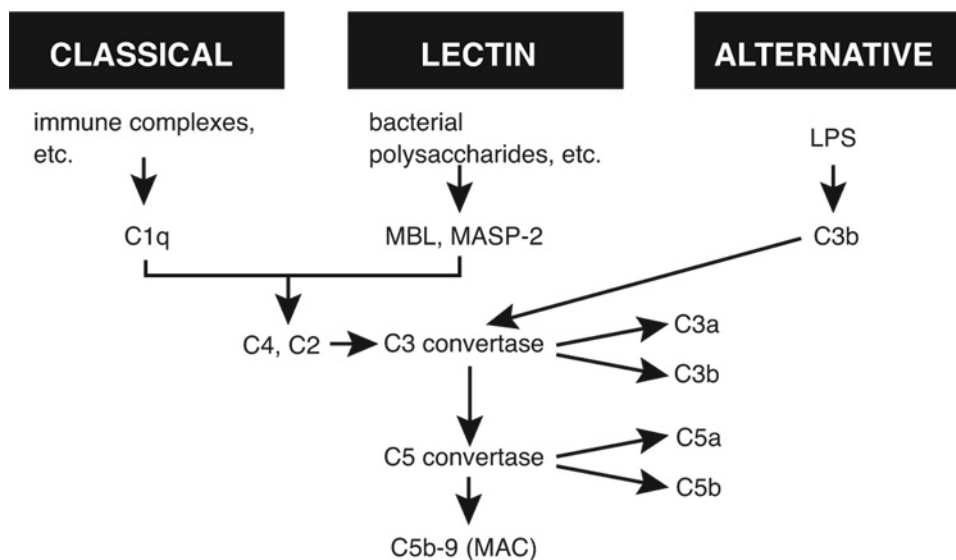


Figure 1: The three pathways of complement activation, collectively resulting in biologically active split products of C3 and C5 (abbreviations: LPS - lipopolysaccharide; MBL – mannose-binding lectin; MASP-2 - MBL-associated protein-2; MAC - membrane attack complex) (Ward PA: Sepsis, apoptosis and complement. *Biochem Pharmacol* 2008; 76: 1383-1388).

## 2. MAIN GOALS

With regard to this background, we hypothesized that acute failure of the myocardial pump function is accompanied by significant inflammatory activation, which can play an important role in the development of further clinical complications.

**Our primary aim** was to investigate the immediate effects of a cardiac tamponade on the central and peripheral haemodynamics, with special emphasis on the intestinal circulation. To this end, we designed a clinically relevant, large animal model with which to determine the macro- and microcirculatory changes induced by experimental cardiac tamponade. With this model, we planned to characterize the major components of the pro-inflammatory profile of the post-tamponade phase, in association with the changes in overall haemodynamics. We assumed that a better understanding of the elements of this mechanism may lead to new prospects for interventions designed to dampen or reverse the secondary, detrimental consequences of acute heart failure.

**Our secondary aim** was to investigate the inhibitory effects of the complement C5a antagonist acetyl-peptide-A (AcPepA), a synthetic, antisense, 17 amino acid peptide acetylated at the N-terminal alanine, on the early circulatory and inflammatory changes in our animal model of cardiac tamponade. It is widely accepted that inflammatory activation plays a decisive role in these conditions, although the potential of ‘anti-inflammatory’ compounds to prevent or cure low perfusion-induced *in vivo* processes is very limited. With this background, we hypothesized that the early inhibition of C5a might well reduce the adverse haemodynamic and inflammatory consequences after the relief of the cardiac tamponade.

### 3. MATERIALS AND METHODS

The experiments were carried out in strict adherence to the NIH guidelines for the use of experimental animals and the study was approved by the Ethical Committee for the Protection of Animals in Scientific Research at the University of Szeged (V./148/2013).

#### 3.1. Instrumentation of animals in Studies I and II

Inbred Vietnamese minipigs of both sexes (weighing  $24 \pm 3$  kg) were fasted for 12 h preoperatively, but received water *ad libitum*. Anaesthesia was induced with a mixture of ketamine ( $20 \text{ mg kg}^{-1}$ ) and xylazine ( $2 \text{ mg kg}^{-1} \text{ im}$ ) and maintained with a continuous infusion of propofol ( $50 \mu\text{L min}^{-1} \text{ kg}^{-1} \text{ iv}$ ;  $6 \text{ mg kg}^{-1} \text{ h}^{-1}$ ) supplemented with ketamine ( $5 \text{ mg kg}^{-1}$ ) - xylazine ( $0.5 \text{ mg kg}^{-1}$ ) mixture *im* before thoracotomy and after 90 min of the observation period. After endotracheal intubation, the animals were mechanically ventilated with the tidal volume set at  $9 \pm 2 \text{ mL kg}^{-1}$ , and the respiratory rate was adjusted to maintain the end-tidal pressure of  $\text{CO}_2$  and arterial  $\text{pCO}_2$  in the range 35-45 mmHg. Positive end-expiratory pressure was not applied during the cardiac tamponade.

The animals were placed in a supine position on a heating pad for maintenance of the body temperature between 36 and 37 °C, and received an infusion of Ringer's lactate at a rate of  $10 \text{ mL kg}^{-1} \text{ h}^{-1}$  during the experiments. The right jugular vein was cannulated (7 F; Edwards Lifesciences LLC, Irvine, U.S.A.) for the measurement of central venous pressure (CVP) and for fluid administration. The right femoral artery and vein were dissected and a thermodilution catheter (PULSION Medical Systems SE, Munich, Germany) was placed in the femoral artery for the measurement of mean arterial pressure (MAP) and CO by a transpulmonary thermodilution method (PiCCO; PULSION Medical Systems SE, Munich, Germany). A pulmonary artery catheter (PV2057 VoLEF Catheter; PULSION Medical Systems SE, Munich, Germany) was inserted via the right femoral vein into the pulmonary artery by tracing the pressure signals.

After a midline abdominal incision, the root of the superior mesenteric artery (SMA) was dissected free. An ultrasonic flow probe (Transonic Systems Inc., Ithaca, NY, U.S.A.) was placed around the exposed SMA to measure the mesenteric blood flow.

Left lateral thoracotomy was performed and a cannula was fixed into the pericardial cavity in all groups. In all protocols, the animals were monitored continuously and a period of 30 min was allowed for recovery from surgery. At the end of the experiments, small intestinal tissue biopsies were taken for biochemical and immunohistochemical analysis and the animals were killed with an overdose of sodium pentobarbital.



### **3.2. Haemodynamic measurements in Studies I and II**

CVP and blood flow signals were monitored continuously and registered with a computerized data-acquisition system (SPELL Haemosys; Experimetria Ltd., Budapest, Hungary). For further haemodynamic monitoring, we used a combination of PiCCO Plus V 5.2.2 and VoLEF V 1.0 (PULSION Medical Systems SE, Munich, Germany) monitors (Végh *et al.* 2009). The MAP, CO, HR, GEDV and extravascular lung water index (EVLWI) were measured with the PiCCO Plus monitoring system, while the PiCCO–VoLEF monitor system was applied to measure the pulmonary arterial pressure (PAP), and calculate RHEDV and LHEDV, systemic vascular resistance index (SVRI) and pulmonary vascular resistance index (PVRI). All haemodynamic parameters were indexed for body surface area or body weight. These techniques were previously used and validated in an accompanying study (Molnár *et al.* 2011). A detailed description of the transpulmonary thermodilution and volumetric analysis is provided elsewhere (Phillips *et al.* 2009).

### **3.3. Evaluation of intestinal microcirculation: pCO<sub>2</sub> gap measurements**

A difference between local tissue and arterial pCO<sub>2</sub> (paCO<sub>2</sub>) levels is a sensitive parameter with which to evaluate the effectiveness of therapy aimed at counteracting a microcirculatory dysfunction in the gastrointestinal tract (Kolkman *et al.* 2000). A silastic balloon-free tonometric probe (Tonosoft Medical Technical and R&D Ltd., Hungary) was introduced through a small enterotomy into the intestinal lumen (Boda *et al.* 2006) to monitor intramucosal pCO<sub>2</sub> levels by capnometry. For calculation of the pCO<sub>2</sub> gap values, simultaneously taken paCO<sub>2</sub> levels were subtracted from the tonometric pCO<sub>2</sub> levels. Arterial and venous blood samples were taken at the baseline and after every hour, and blood-gas parameters were measured with a blood-gas analyser (Cobas b121, Roche, Austria).

### **3.4. Biochemical measurements**

Five-mL blood samples were drawn from the jugular vein into chilled polypropylene tubes containing EDTA (1 mg mL<sup>-1</sup>) as the scheduled experimental protocol. The blood samples were centrifuged at 1200g for 10 min at 4 °C. The plasma samples were next collected and stored at -70 °C until assay.

#### *Big-endothelin measurements in plasma*

Blood samples were collected into chilled polypropylene tubes containing EDTA. Plasma levels of big-ET, a 38 amino acid stable precursor protein of ET-1, were measured with a commercially available kit (Biochemica Hungaria Kft., Budapest, Hungary).

#### *Tumour necrosis factor-alfa measurements in plasma*

The plasma concentration of tumour necrosis factor-alfa (TNF- $\alpha$ ) was measured with a commercially available porcine ELISA kit (Biochemica Hungaria Kft., Budapest, Hungary).

#### *High-mobility group box protein-1 measurements in plasma*

The plasma concentration of HMGB-1 was measured with a commercially available HMGB-1 ELISA kit (Shino-Test Corporation, Kanagawa, Japan).

#### *Histamine measurements in plasma*

Plasma histamine concentrations were determined by a commercially available enzyme-linked immunoassay (Quantikine ultrasensitive EIA kit for histamine; Biomedica Hungaria Kft, Budapest, Hungary).

#### *Plasma nitrite/nitrate level measurements*

The levels of plasma nitrite/nitrate (NO<sub>x</sub>), stable end-products of NO, were measured by means of the Griess reaction. The assay depends on the enzymatic reduction of nitrate to nitrite, which is then converted into a coloured azo compound detected spectrophotometrically at 540 nm (Moshage *et al.* 1995).

#### *Measurement of whole-blood superoxide production*

For the whole-blood superoxide production measurements, the chemiluminometric method of Zimmermann *et al.* was used (Zimmermann *et al.* 1991). During the measurements, 10  $\mu$ L of whole blood was added to 1 mL of Hanks solution and the mixture was kept at 37 °C until assay. The chemiluminometric response was measured with a Lumat LB9507 luminometer (Berthold, Vienna, Austria) during 30 min after the addition of 100  $\mu$ L of lucigenin.

#### *Plasma troponin-T level measurements*

Cardiac troponin-T levels in plasma samples were measured by highly sensitive electrochemiluminescent immunoassay (ECLIA; Elecsys 2010, Roche Diagnostics GmbH,

Mannheim, Germany). The analytical sensitivity was 5 ng L<sup>-1</sup>, and the intra- and interassay variations in the measured concentration range were 3.2 and 6.2 CV%, respectively.

#### *Myeloperoxidase activity measurement*

The activity of myeloperoxidase (MPO), as a marker of tissue PMN leukocyte infiltration, was measured via intestinal biopsies (Kuebler *et al.* 1996). Briefly, the sample was homogenized with Tris-HCl buffer (0.1 M, pH 7.4) containing 0.1 mM phenylmethylsulfonyl fluoride to block tissue proteases, and then centrifuged at 4 °C for 20 min at 2000g. The MPO activities of the samples were measured at 450 nm (UV-1601 spectrophotometer, Shimadzu, Japan) and the data were referred to the protein content.

#### *Immunohistochemistry*

The presence of a complement C3 deposit in the small intestinal mucosa was detected by immunohistochemistry (IHC) (Girardi *et al.* 2003) on formalin-fixed, paraffin-embedded small intestinal sections, using rabbit polyclonal anti-complement fragment C3c primary antibody (Bioss Inc, Woburn, MA, USA). The sections were deparaffinized for 5 min, which was followed by antigen retrieval with citrate buffer for 20 min. The activity of endogenous peroxidases was blocked with 5% H<sub>2</sub>O<sub>2</sub> for 10 min. The non-specific interactions were inhibited during the next 30 min of incubation. The primary antibody was diluted 1:500 in antibody diluent for IHC (BD Pharmingen, San Diego, CA, USA). After washing of the sections, the secondary antibody was diluted 1:500 and the samples were incubated for 8 min. Sections were counterstained with 3,3'-diaminobenzidine and haematoxylin. The entire IHC investigation was carried out with an automatic Leica Bond-max IHC instrument (Leica Microsystems, Tokyo, Japan). For quantitative analysis, immunostained sections were examined under a light microscope. The coded sections were analysed by an independent histopathology specialist. The numbers of capillaries positive for complement fragment C3c were assessed at a magnification of 400x.

### **3.5. Experimental protocols**

*In Study I*, the animals were randomly divided into two experimental groups. Group 1 (n=7) served as sham-operated control, with the same time-frame and sampling as in group 2. A pericardial tamponade (group 2; n=8) was induced for 60 min by the intrapericardial administration of colloid solution (60-90 mL hydroxyethyl starch 6%; Fresenius Kabi Deutschland GmbH, Homburg, Germany), while the MAP was kept in the interval 40-45

mmHg. After this period, the fluid was removed from the pericardial sac and the animals were monitored for 180 min post-tamponade.

Peripheral blood samples were taken at baseline, and then after 75 min, after 150 min and at the end of the observation period (240 min) to detect the levels of vasoactive and inflammatory mediators (big-ET, histamine, NO<sub>x</sub>, troponin-T, HMGB-1 and whole-blood superoxide production). Small intestinal tissue biopsies were taken at the end of the experiments for MPO activity measurements and IHC analysis of the complement C3 deposit.

*In Study II*, the animals were randomly allocated into one or other of three experimental groups with the same time-frame and sampling as in Study I. Group 1 (n=6) served as sham-operated control, while in groups 2 (n=7) and 3 (n=6) cardiac tamponade was induced. Group 3 was treated with C5a antagonist AcPepA (a single administration of 4 mg kg<sup>-1</sup> in 5 mL saline *iv* into the jugular vein in a 5-min infusion) after 45 min of cardiac tamponade. AcPepA (ASGAPAPGPAGPLRPMF) containing an acetylated N-terminal alanine was synthesized and purified (> 95% purity) by Biologica Co. Ltd. (Nagoya, Japan). The peptide was dissolved in saline and used in a concentration of 2 mg mL<sup>-1</sup> as reported previously (Okada *et al.* 2011). Vehicle (saline) administration was applied in groups 1 and 2 by the same protocol. The beginning of tamponade is denoted as 0 min. Blood and tissue sampling were performed during the 240 min observation period as described in Study I.

### **3.6. Statistical analysis**

Data analysis was performed with a statistical software package (SigmaStat for Windows, Jandel Scientific, Erkrath, Germany). The distribution of our experimental data was analysed by the Kolmogorov-Smirnov normality test. Failure of the normality test indicated non-parametric distribution of the data. In Studies I-II, non-parametric methods were used. Friedman repeated measures analysis of variance on ranks was applied within the groups. Time-dependent differences from the baseline (time 0) for each group were assessed by Dunn's method, and differences between groups were analysed with the Mann-Whitney test (in Study I) or with Kruskal-Wallis one-way analysis of variance on ranks, followed by Dunn's method for pairwise multiple comparison (in Study II). In the Figures and Tables, median values and the 25<sup>th</sup> (lower whisker) and 75<sup>th</sup> (upper whisker) percentiles are given. \**p*<0.05 within groups *vs* baseline values, <sup>x</sup>*p*<0.05 between groups *vs* sham-operated group values, <sup>#</sup>*p*<0.05 between AcPepA-treated group *vs* cardiac tamponade group.

## 4. RESULTS

### 4.1. Study I – Haemodynamic and inflammatory responses after cardiac tamponade

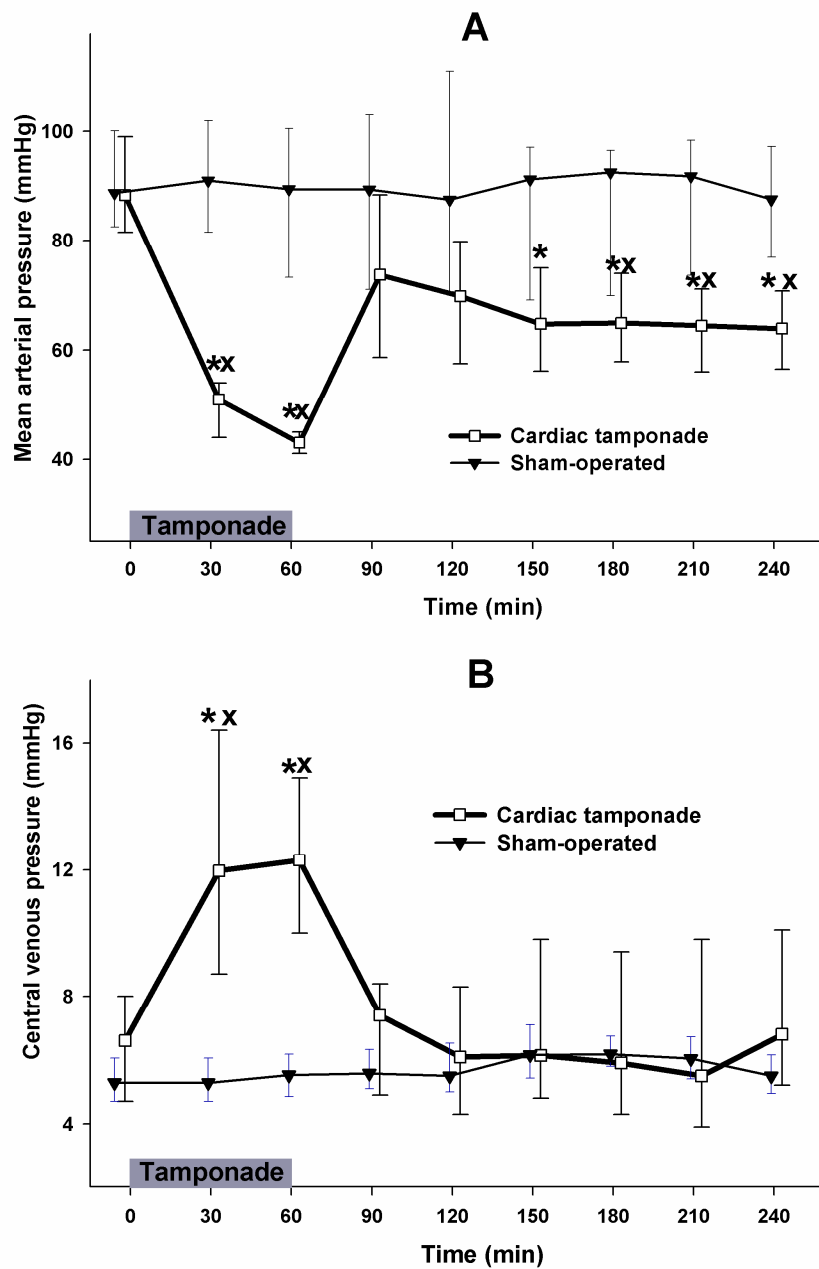
#### 4.1.1. Changes in haemodynamics

In the sham-operated group, there were no significant haemodynamic changes as compared with the baseline values, and the mediator levels did not change significantly during the observation period.

The MAP was maintained in the interval 40-45 mmHg during the tamponade for 60 min (Figure 2A) by the infusion of colloid fluid into the pericardial sac, which resulted in a significant, approximately 60% decline in CO in the group undergoing cardiac tamponade. The SVRI and HR were increased significantly (by 32% and 66%, respectively; Table 1). After relief of the tamponade, the MAP was significantly lower in the cardiac tamponade group as compared with the control group, while the CO and HR returned to the baseline despite the reduced MAP.

**Table 1.** Effects of cardiac tamponade on the CO, HR and SVRI. Values are medians (25th percentile; 75th percentile). \*  $p < 0.05$  within group; <sup>X</sup>  $p < 0.05$  between groups vs control group;

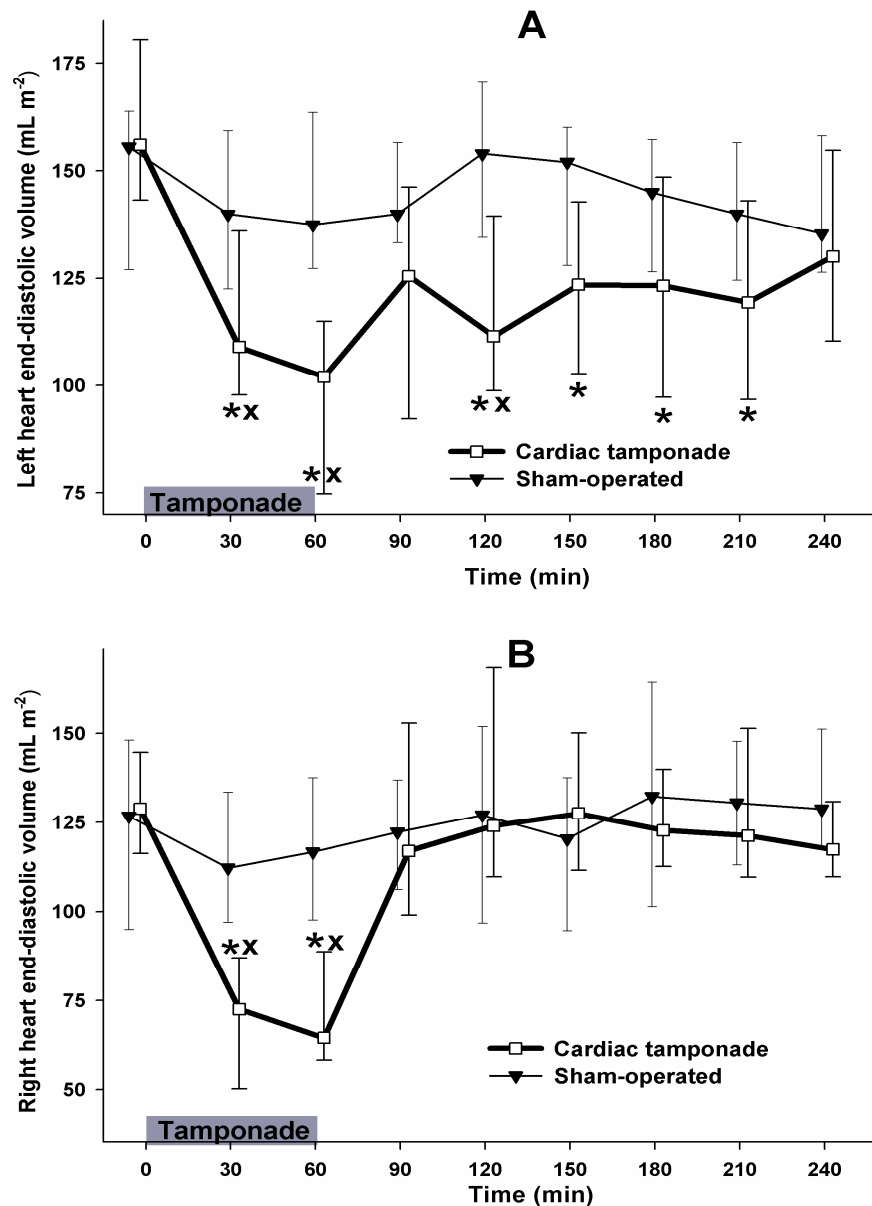
|  | -5 min               | 30 min                              | 60 min                              | 90 min               | 120 min                           | 180 min                           | 240 min              |
|--|----------------------|-------------------------------------|-------------------------------------|----------------------|-----------------------------------|-----------------------------------|----------------------|
| CO [L min <sup>-1</sup> m <sup>-2</sup> ]                        |                      |                                     |                                     |                      |                                   |                                   |                      |
| Sham-operated  | 2.60<br>(2.39; 2.97) | 2.73<br>(2.25; 3.34)                | 2.74<br>(2.30; 3.14)                | 2.85<br>(2.60; 3.28) | 2.94<br>(2.65; 3.30)              | 2.85<br>(2.51; 3.35)              | 2.82<br>(2.62; 3.33) |
| Cardiac tamponade  | 2.82<br>(2.46; 3.01) | 1.36 * <sup>X</sup><br>(0.95; 1.89) | 1.19 * <sup>X</sup><br>(1.03; 1.30) | 2.44<br>(2.29; 3.01) | 2.48<br>(2.16; 3.03)              | 2.40<br>(2.13; 2.74)              | 2.40<br>(2.07; 2.79) |
| HR [beat min <sup>-1</sup> ]                                     |                      |                                     |                                     |                      |                                   |                                   |                      |
| Sham-operated  | 124<br>(120; 137)    | 128<br>(118; 134)                   | 125<br>(119; 134)                   | 126<br>(115; 137)    | 127<br>(104; 135)                 | 120<br>(104; 128)                 | 122<br>(106; 126)    |
| Cardiac tamponade  | 113<br>(103; 118)    | 175 * <sup>X</sup><br>(136; 203)    | 188 * <sup>X</sup><br>(173; 206)    | 125<br>(122; 143)    | 118<br>(113; 120)                 | 114<br>(105; 116)                 | 113<br>(98; 122)     |
| SVRI [mmHg mL <sup>-1</sup> min <sup>-1</sup> kg <sup>-1</sup> ] |                      |                                     |                                     |                      |                                   |                                   |                      |
| Sham-operated  | 0.79<br>(0.73; 0.83) | 0.81<br>(0.75; 0.85)                | 0.74<br>(0.69; 0.79)                | 0.68<br>(0.66; 0.77) | 0.64<br>(0.58; 0.68)              | 0.66<br>(0.57; 0.71)              | 0.67<br>(0.61; 0.75) |
| Cardiac tamponade  | 0.78<br>(0.71; 0.92) | 0.91<br>(0.80; 0.94)                | 1.03 *<br>(0.84; 1.06)              | 0.83<br>(0.72; 0.85) | 0.83 <sup>X</sup><br>(0.69; 0.93) | 0.82 <sup>X</sup><br>(0.73; 0.91) | 0.79<br>(0.71; 0.91) |



**Figure 2.** Changes in mean arterial pressure (A) and central venous pressure (B) in the sham-operated (triangles with thin continuous line) and cardiac tamponade (squares with thick solid line) groups. The box indicates the duration of the cardiac tamponade.

The decline in the venous return during the tamponade was evidenced by the increased CVP (Figure 2B). This process was accompanied by decreases in RHEDV and LHEDV (Figure 3AB). After relief of the tamponade, the CVP and RHEDV normalized, but the LHEDV did not reach the baseline value in the tamponade group; it remained significantly

lower as compared with the sham-operated group (Figures 3AB). These changes demonstrate the long-lasting impairment of the LV function following the cardiac tamponade.



**Figure 3.** Changes in right heart end-diastolic volume (A) and left heart end-diastolic volume (B) in the sham-operated (triangles with thin continuous line) and cardiac tamponade (squares with thick solid line) groups. The box indicates the duration of the cardiac tamponade.

The cardiac tamponade resulted in a significant, transient decrease in PAP and a 2-fold elevation of the PVRI (Table 2). In the post-tamponade period, further long-lasting significant increases in PVRI and PAP occurred as compared with the sham-operated group, while the EVLWI was significantly elevated at the end of the observation period (Table 2).

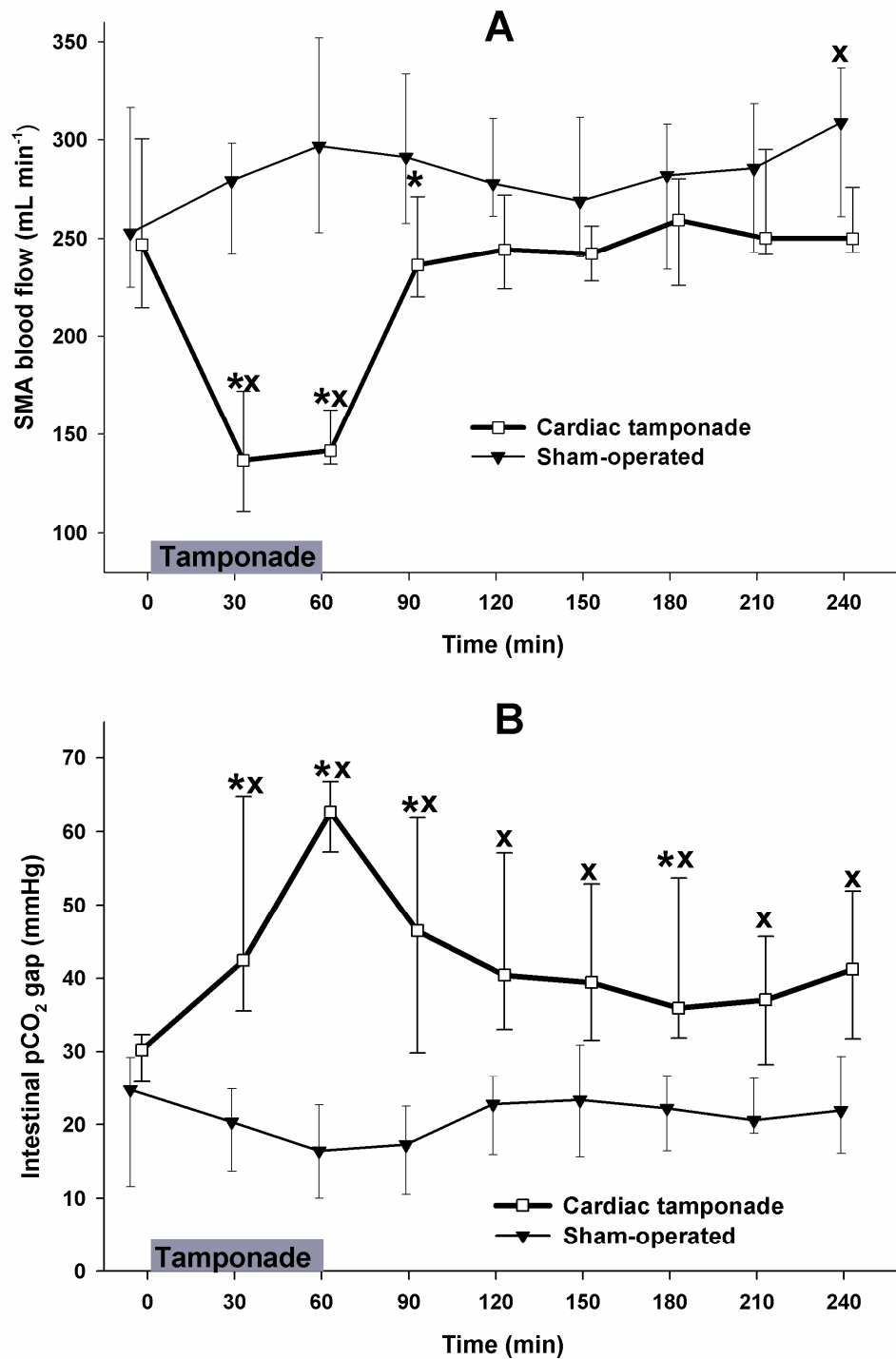
**Table 2.** Effects of cardiac tamponade on PAP, PVRI and EVLWI. Values are medians (25th percentile; 75th percentile). \*  $p < 0.05$  within group; <sup>x</sup>  $p < 0.05$  between groups vs control group;

|  | -5 min               | 30 min                              | 60 min                              | 90 min                            | 120 min                           | 180 min                             | 240 min                            |
|--|----------------------|-------------------------------------|-------------------------------------|-----------------------------------|-----------------------------------|-------------------------------------|------------------------------------|
| PAP [mmHg]   |                      |                                     |                                     |                                   |                                   |                                     |                                    |
| Sham-operated  | 30.6<br>(27.0; 35.0) | 31.1<br>(27.3; 35.4)                | 28.9<br>(27.5; 35.6)                | 31.2<br>(26.0; 36.0)              | 30.3<br>(27.4; 34.8)              | 30.3<br>(26.8; 32.5)                | 30.8<br>(28.3; 33.6)               |
| Cardiac tamponade  | 30.4<br>(27.3; 34.2) | 24.3 * <sup>x</sup><br>(21.1; 27.8) | 30.0<br>(19.2; 35.6)                | 38.4<br>(30.0; 41.3)              | 36.5<br>(34.4; 42.7)              | 41.2 * <sup>x</sup><br>(35.3; 43.2) | 37.9<br>(31.9; 42.8)               |
| PVRI [mmHg mL <sup>-1</sup> min <sup>-1</sup> kg <sup>-1</sup> ] |                      |                                     |                                     |                                   |                                   |                                     |                                    |
| Sham-operated  | 0.37<br>(0.31; 0.39) | 0.35<br>(0.30; 0.43)                | 0.31<br>(0.29; 0.35)                | 0.29<br>(0.25; 0.35)              | 0.29<br>(0.25; 0.32)              | 0.32<br>(0.24; 0.36)                | 0.31<br>(0.23; 0.38)               |
| Cardiac tamponade  | 0.41<br>(0.26; 0.50) | 0.75 * <sup>x</sup><br>(0.66; 0.98) | 0.88 * <sup>x</sup><br>(0.81; 0.95) | 0.48 <sup>x</sup><br>(0.42; 0.53) | 0.51 <sup>x</sup><br>(0.44; 0.63) | 0.54 <sup>x</sup><br>(0.49; 0.60)   | 0.49 <sup>x</sup><br>(0.47; 0.54)  |
| EVLV [mL kg <sup>-1</sup> ]                                      |                      |                                     |                                     |                                   |                                   |                                     |                                    |
| Sham-operated  | 8.0<br>(7.8; 9.0)    | 8.2<br>(8.0; 9.0)                   | 8.5<br>(8.0; 9.0)                   | 8.5<br>(8.0; 9.1)                 | 8.5<br>(8.0; 9.1)                 | 8.4<br>(8.0; 9.0)                   | 8.4<br>(8.0; 9.0)                  |
| Cardiac tamponade  | 8.0<br>(8.0; 9.0)    | 8.0<br>(7.5; 8.8)                   | 8.0<br>(7.3; 8.9)                   | 8.5<br>(8.0; 10.8)                | 9.0<br>(8.3; 10.8)                | 9.5 *<br>(8.5; 10.0)                | 10.0 * <sup>x</sup><br>(9.0; 11.0) |

The significant decrease in SMA blood flow during the tamponade indicated a deteriorated mesenteric circulation. After the removal of the pericardiac fluid, the SMA flow returned to the control values (Figure 4A).

The pCO<sub>2</sub> gap, the difference between the local tissue and the arterial pCO<sub>2</sub>, is a reliable index of local tissue perfusion. The pCO<sub>2</sub> gap increased significantly during the tamponade, while relief of the tamponade resulted in a significantly lower gap, though the values remained significantly higher than that for the sham-operated control group throughout the post-tamponade period (Figure 4B).

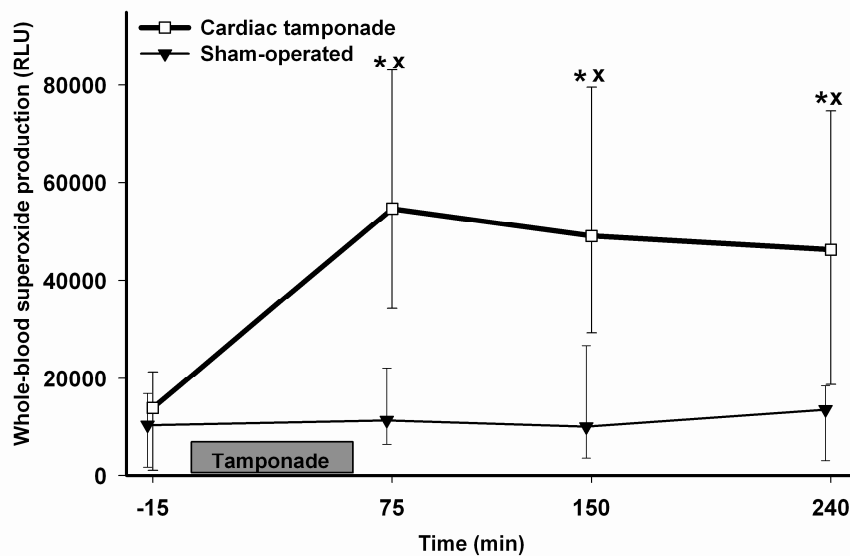




**Figure 4.** Changes in superior mesenteric artery (SMA) blood flow (**A**) and intestinal  $\text{pCO}_2$  gap (**B**) in the sham-operated (triangles with thin continuous line) and cardiac tamponade (squares with thick solid line) groups. The box indicates the duration of the cardiac tamponade.

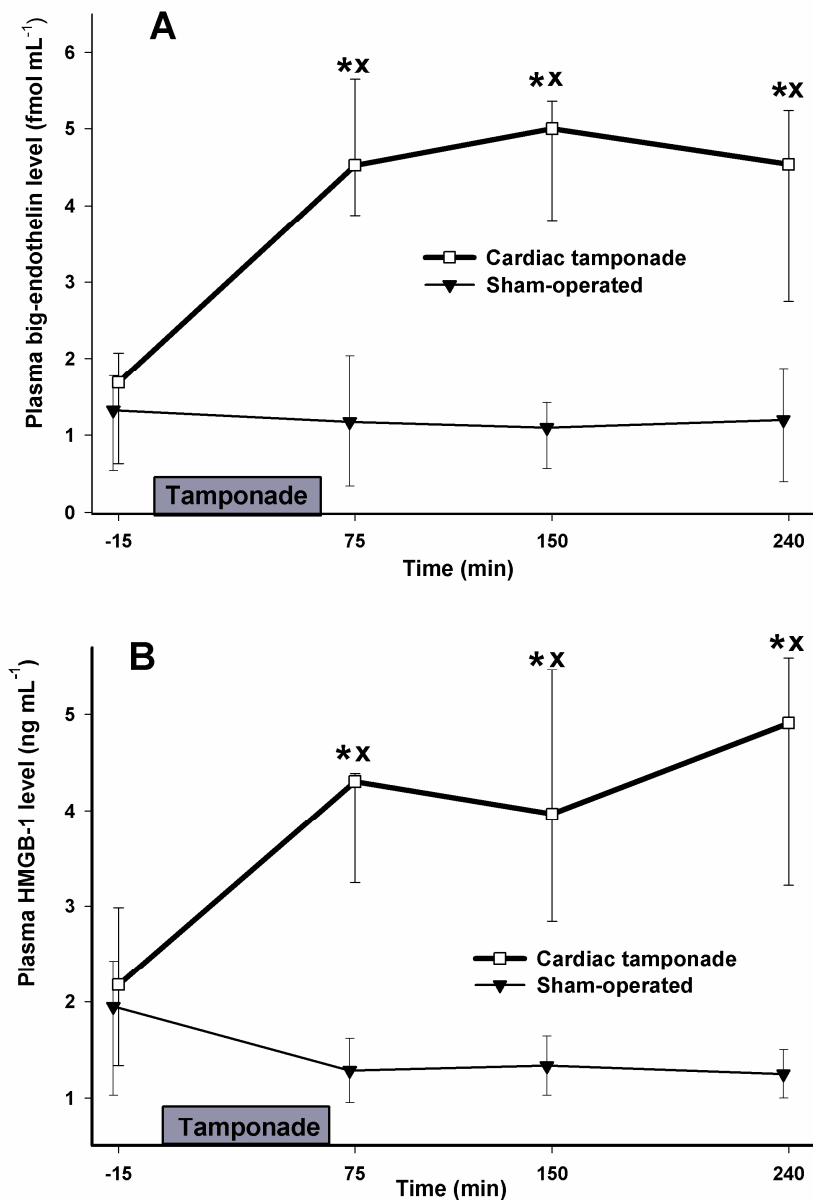
#### 4.1.2. Changes in biochemical parameters

In the cardiac tamponade group, increased superoxide radical production was observed at the beginning of the post-tamponade phase (Figure 5). In parallel, the plasma histamine levels were increased significantly by 75 and 240 min of the observation period:  $M=16.2$  nM ( $p_{25}=15.5$ ;  $p_{75}=16.6$ ) and  $M=10.3$  nM ( $p_{25}=9.1$ ;  $p_{75}=12.8$ ), respectively, *vs* the baseline ( $M=7.5$ ;  $p_{25}=6.4$ ;  $p_{75}=8.8$ ), or *vs* the corresponding value for the sham-operated group ( $M=7.4$  nM;  $p_{25}=5.7$ ;  $p_{75}=9.4$ ).



**Figure 5.** Changes in whole-blood superoxide production in the sham-operated (triangles with thin continuous line) and cardiac tamponade (squares with thick solid line) groups. The box indicates the duration of the cardiac tamponade.

The  $\text{NO}_x$  concentration in the plasma allows an estimate of the changes in NO production. The consequence of the cardiac tamponade in this respect was a slight, but statistically significant increase in  $\text{NO}_x$  level at the end of the post-tamponade period as compared with the baseline level and with that for the sham-operated group (data not shown).



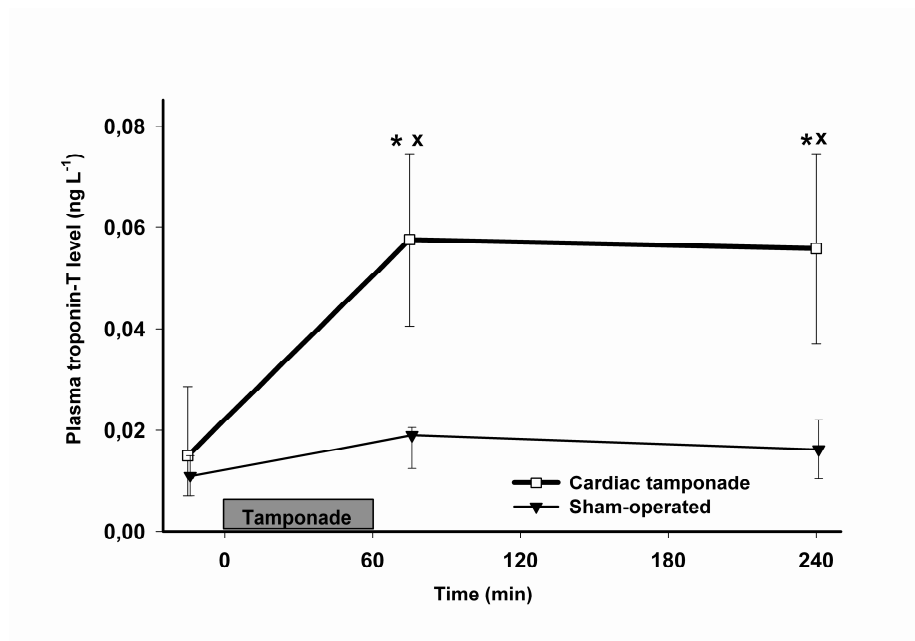
**Figure 6.** Changes in plasma big-endothelin (ET) concentration (A) and high-mobility group box protein-1 (HMGB-1) level (B) in the sham-operated (triangles with thin continuous line) and cardiac tamponade (squares with thick solid line) groups. The box indicates the duration of the cardiac tamponade.

Big-ET is a stable precursor of ET-1 with a longer half-life. The plasma big-ET level increased significantly (4-5-fold) in response to the cardiac tamponade (Figure 6A).

HMGB-1 is an effective signal for leukocyte activation, which causes an escalation of the inflammatory process. The plasma level of HMGB-1 was elevated significantly after the compression of the heart (Figure 6B).

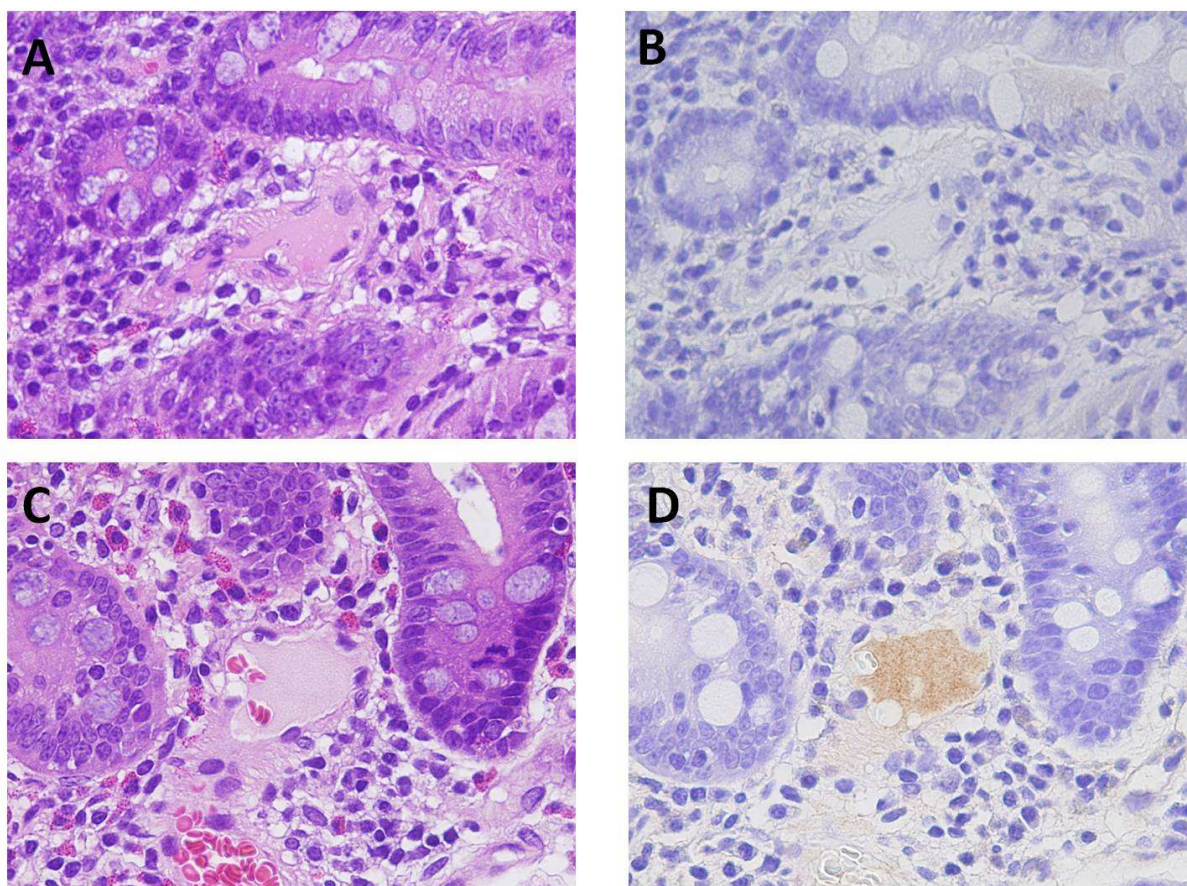
The rate of PMN leukocyte accumulation was determined through measurement of the MPO activity of the intestinal tissue samples taken at the end of the experiments. The level of MPO activity was significantly higher in the small intestinal tissue samples of the cardiac tamponade group, indicating the increased accumulation of PMN leukocytes (cardiac tamponade:  $M=5.37$  ( $p25=4.8$ ;  $p75=6.22$ ;  $U \text{ mg protein}^{-1}$ ) vs sham-operated:  $M=2.84$  ( $p25=2.19$ ;  $p75=3.22$ )).

The troponin-T level in the plasma allows an estimate of the cardiomyocyte damage. The mean concentrations of troponin-T were significantly increased after the compression of the heart and at the end of the post-tamponade period as compared with the baseline and with those for the sham-operated group (Figure 7).



**Figure 7.** Changes in plasma troponin-T level in the sham-operated (triangles with thin continuous line) and cardiac tamponade (squares with thick solid line) groups. The box indicates the duration of the cardiac tamponade.

Activation of the complement cascade was evaluated by the presence of a complement C3 deposit in the small intestinal mucosa with the IHC method (Figure 8). In each field of view of the slides, the number of capillaries showing C3 deposit positivity was counted. The number of C3 deposits was significantly higher in the tamponade group than in the sham-operated group ( $M=3$ ;  $p25=1$ ;  $p75=4.5$  vs  $M=0$ ;  $p25=0$ ;  $p75=1$  deposits/field of view).



**Figure 8.** Changes in complement C3 deposition as demonstrated by IHC analyses in the small intestinal mucosal biopsies taken at the end of the experiments: H&E staining in the sham-operated (A) and cardiac tamponade (C) groups; extensive C3 deposition with IHC staining in the cardiac tamponade (D) group as compared with the sham-operated (B) group.

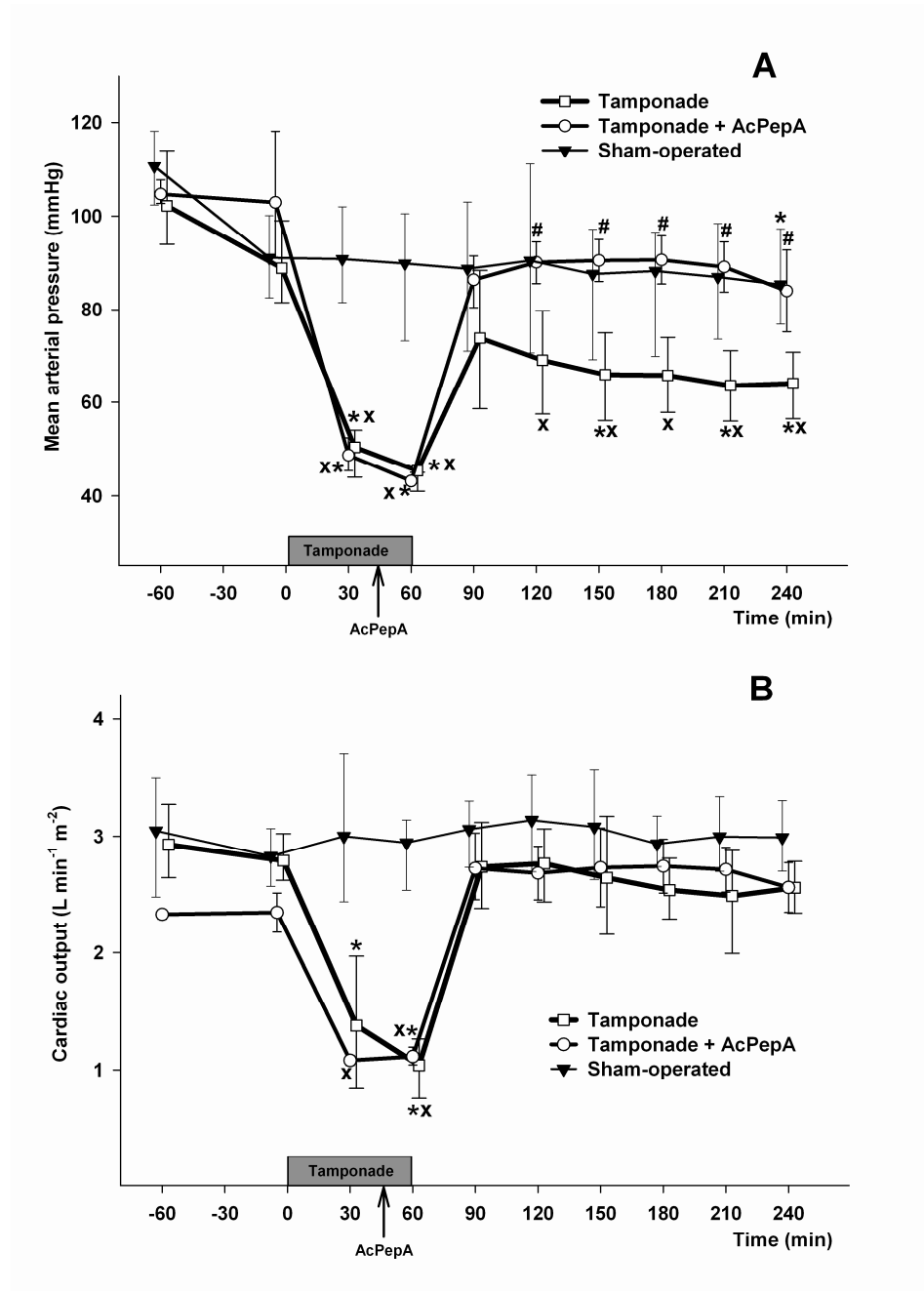
## 4.2. Study II – Cardiac effects of complement C5a antagonist treatment

### 4.2.1. Changes in cardiac and pulmonary haemodynamics

In the sham-operated group, there were no significant haemodynamic changes as compared with the baseline values, and the mediator levels did not change significantly during the observation period.

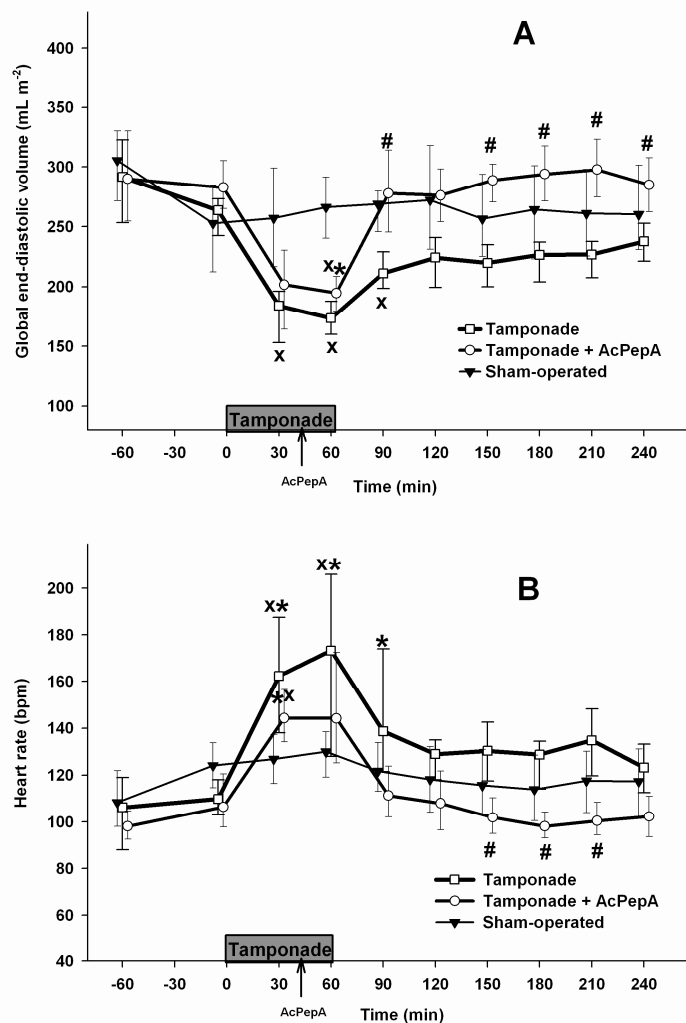
MAP was kept in the interval 40-45 mmHg during cardiac tamponade for 60 min (Figure 9A) by the infusion of colloid fluid into the pericardial sac, and this resulted in a significant 65% decline in CO in both groups undergoing cardiac tamponade (Figure 9B) and a significant increase in HR (Figure 10B). After relief of the tamponade, the MAP was significantly lower in the non-treated cardiac tamponade group as compared with the control group, while the CO and HR returned to the baseline values despite the reduced MAP.

Administration of AcPepA after 45 min of the cardiac tamponade resulted in an elevation of the MAP, did not influence the CO, and caused a significant decrease in the HR as compared with the untreated cardiac tamponade group in the post-tamponade period (Figures 9A-B and 10B).



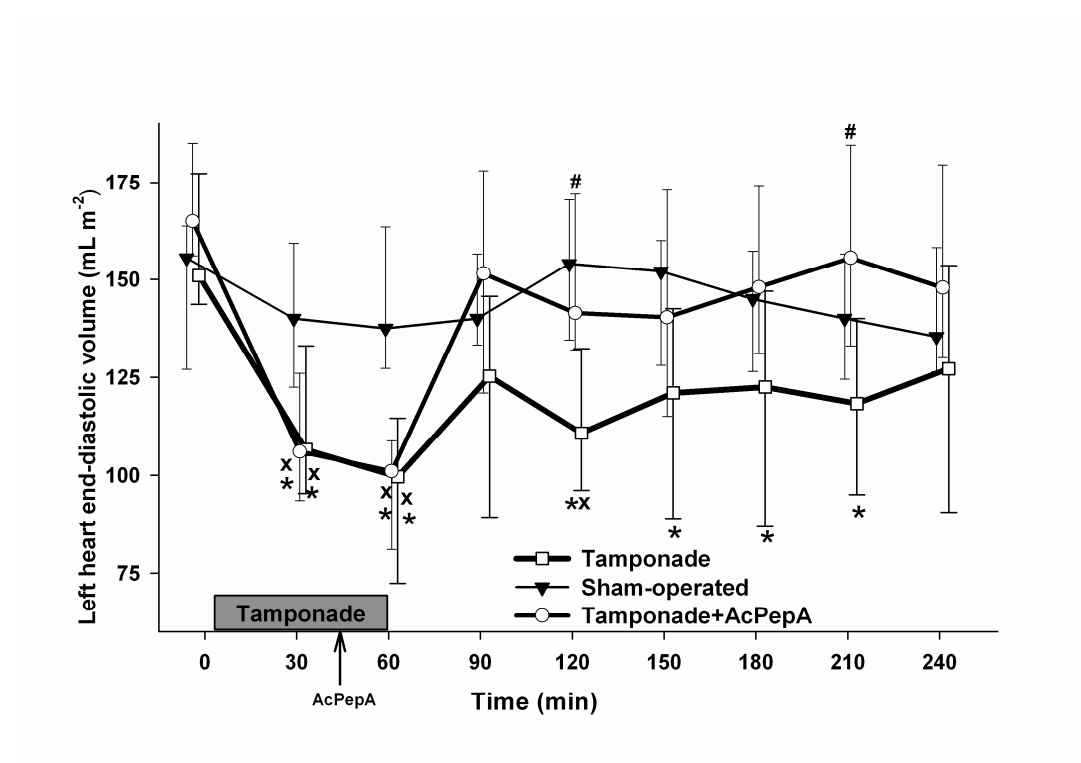
**Figure 9.** Changes in mean arterial pressure (A) and cardiac output (B) in the sham-operated (solid triangles with continuous line), cardiac tamponade (empty squares with solid line) and AcPepA-treated (empty circles with solid line) groups. The box indicates the duration of the cardiac tamponade, and the arrow indicates the time of treatment with AcPepA.

The decline in the venous return during the tamponade was evidenced by the increased CVP (Figure 2B). This process was accompanied by a decrease in GEDV (Figure 10A). The GEDV did not reach the baseline values in the non-treated cardiac tamponade group (Figure 10A) after relief of the tamponade. These changes demonstrate the long-lasting impairment of the venous return flow following the cardiac tamponade. After AcPepA administration, we observed significant increases in the preload parameters in the post-tamponade period: the CVP was set to a significantly higher level (Annex II, Figure 2A) and the GEDV demonstrated the increased returning blood flow (Figure 10A).



**Figure 10.** Changes in global end-diastolic volume (A) and heart rate (B) in the sham-operated (solid triangles with continuous line), cardiac tamponade (empty squares with solid line) and AcPepA-treated (empty circles with solid line) groups. The box indicates the duration of the cardiac tamponade, and the arrow indicates the time of treatment with AcPepA.

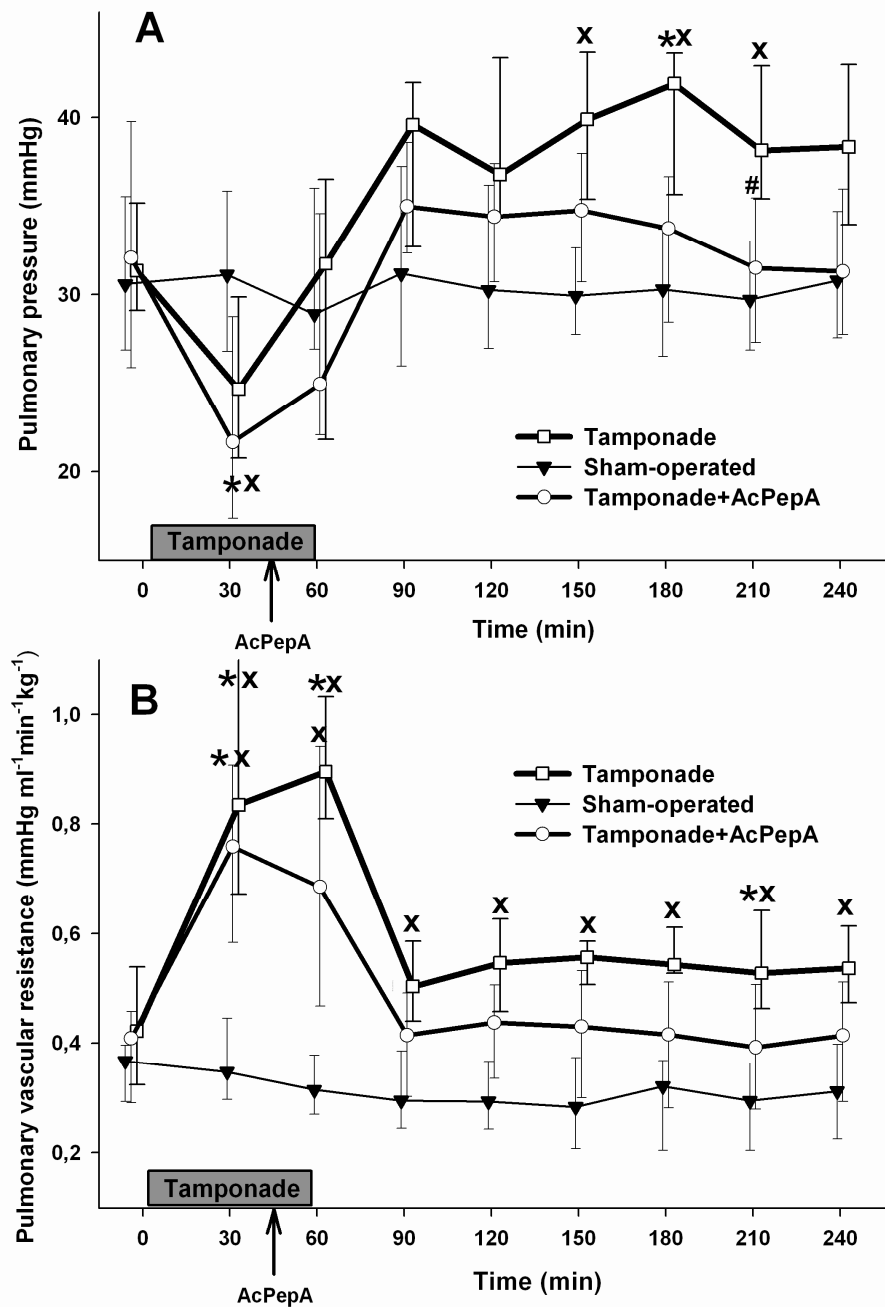
The cardiac tamponade caused a permanent impairment of the LV function, as demonstrated by the long-lasting decrease in LHEDV (Figure 11). Administration of AcPepA after 45 min of the cardiac tamponade normalized the LHEDV as compared with the untreated cardiac tamponade group in the post-tamponade period. In contrast with the LHEDV, the RHEDV returned to the baseline level after the relief of the tamponade (Figure 3B) and it was not influenced by complement C5a antagonist treatment (data not shown).



**Figure 11.** Changes in left heart end-diastolic volume in the sham-operated (solid triangles with continuous line), cardiac tamponade (empty squares with solid line) and AcPepA-treated (empty circles with solid line) groups. The box indicates the duration of the cardiac tamponade, and the arrow indicates the time of treatment with AcPepA.

The cardiac tamponade caused significant changes in the pulmonary circulation in both the tamponade and the post-tamponade periods (Table 2). AcPepA treatment decreased the long-lasting elevations in PAP and PVRI that occurred after the relief of the tamponade as compared with the untreated cardiac tamponade group (Figure 12A,B). However, the elevation caused in EVLWI by the cardiac tamponade was not influenced significantly by this treatment at the end of the observation period (data not shown).

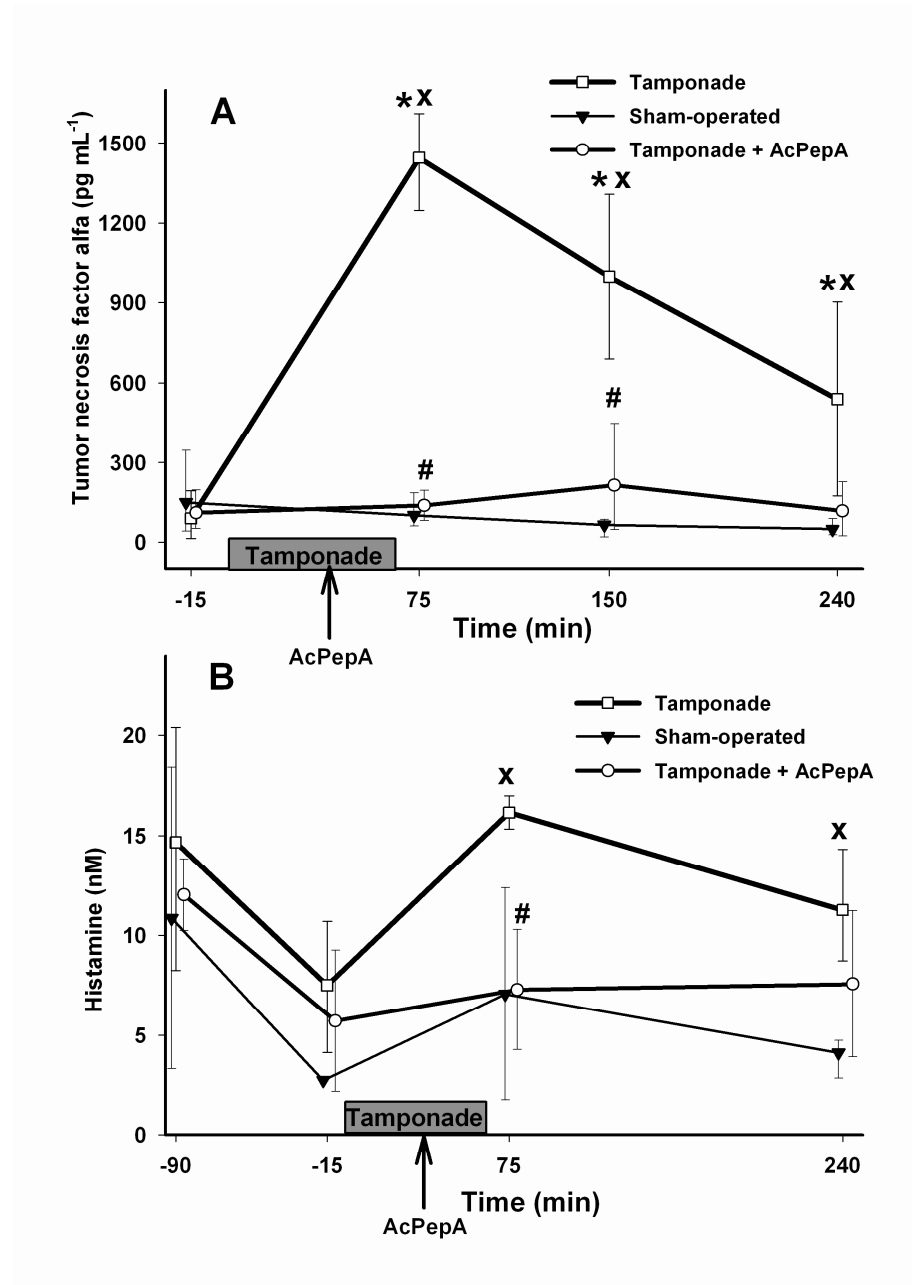




**Figure 12.** Changes in pulmonary arterial pressure (**A**) and pulmonary vascular resistance (**B**) in the sham-operated (solid triangles with continuous line), cardiac tamponade (empty squares with solid line) and AcPepA-treated (empty circles with solid line) groups. The box indicates the duration of the cardiac tamponade, and the arrow indicates the time of treatment with AcPepA.

#### 4.2.2. Changes in biochemical parameters

Change in the TNF- $\alpha$  level is a sensitive and early signal indicative of the inflammatory process. In our pig model, a 10-fold elevation in the plasma level of TNF- $\alpha$  was detected after the compression of the heart; the level then decreased successively until the end of the observation period (Figure 13A).

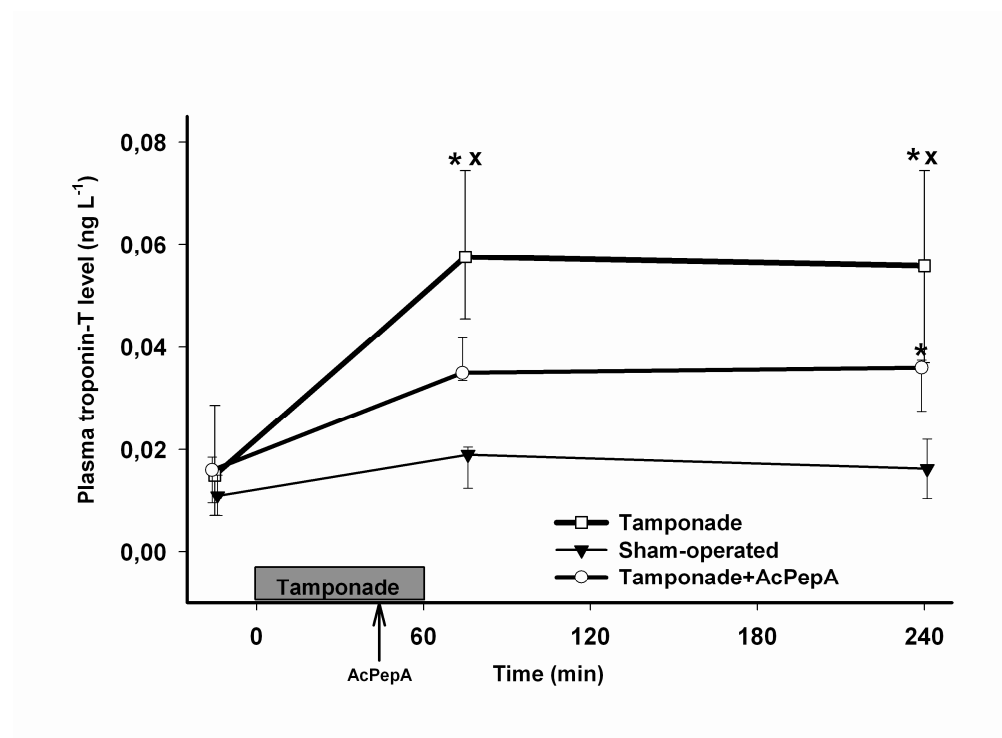


**Figure 13.** Changes in plasma TNF- $\alpha$  (A) and histamine (B) levels in the sham-operated (solid triangles with continuous line), cardiac tamponade (empty squares with solid line) and AcPepA-treated (empty circles with solid line) groups. The box indicates the duration of the cardiac tamponade, and the arrow indicates the time of treatment with AcPepA.

The increased level of histamine release in the plasma suggests the activation of the hypoxia-sensitive inflammatory cells (mast cells and basophil leukocytes). As a result of the cardiac tamponade, the histamine level increased significantly by 15 min of the post-tamponade phase (Figure 13B).

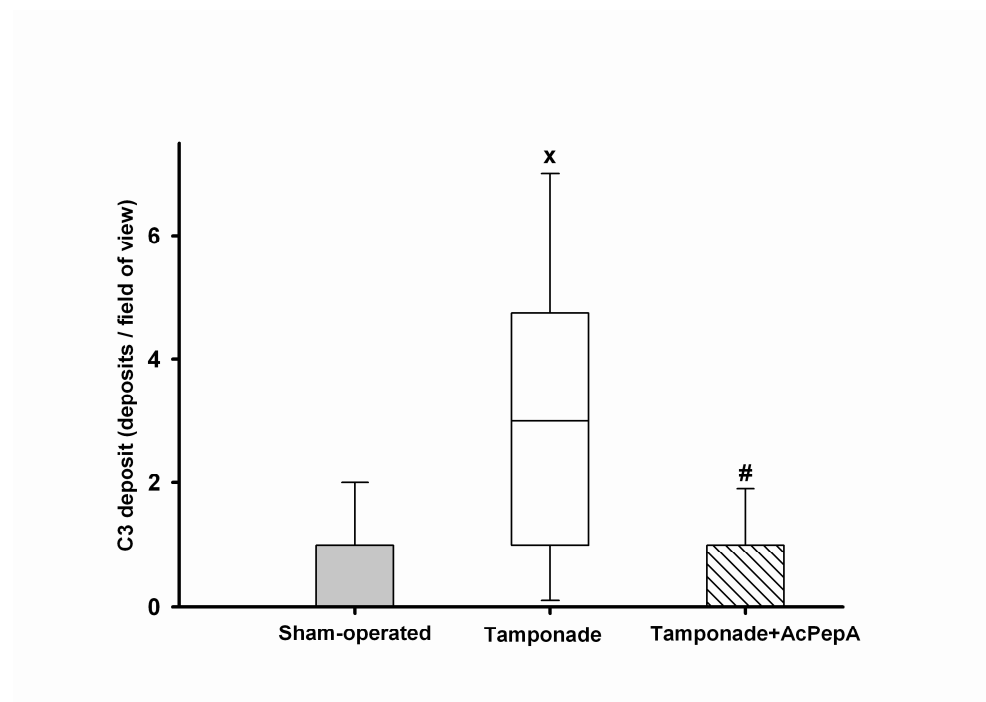
After AcPepA administration, the characteristic biochemical changes following the cardiac tamponade were significantly different. The AcPepA treatment reduced the concentrations of TNF- $\alpha$  and histamine in the plasma after the compression of the heart and in the post-tamponade period as compared with the sham-operated group (Figure 13A,B).

A measurable troponin-T level in the plasma allows an estimate of the ischaemic effect of cardiac tamponade. The cardiac tamponade caused a significant elevation in the mean concentration of troponin-T after the relief of the tamponade and at the end of the post-tamponade period, while the administration of AcPepA resulted in a lower level of troponin-T in the plasma in the post-tamponade phase (Figure 14).



**Figure 14.** Changes in plasma troponin-T level in the sham-operated (solid triangles with continuous line), cardiac tamponade (empty squares with solid line) and AcPepA-treated (empty circles with solid line) groups. The box indicates the duration of the cardiac tamponade, and the arrow indicates the time of treatment with AcPepA.

Activation of the complement cascade was detected via the increased presence of a complement C3 deposit in the tamponade group as compared with the sham-operated group in the small intestinal mucosa with the IHC method. However, the number of capillaries displaying C3 deposit positivity was lower in the AcPepA-treated tamponade group than in the non-AcPepA-treated group (Figure 15).



**Figure 15.** Changes in complement C3 deposition demonstrated by immunohistochemical analyses in the small intestinal mucosal biopsies taken at the end of the experiments.

## 5. DISCUSSION

### 5.1. Study I

Pericardial tamponade is accompanied by high mortality and postoperative complication rates, even in the event of adequate treatment. In cases of blunt chest trauma, the mortality may exceed 80% (Brathwaite *et al.* 1990), while a 22% mortality rate was reported after elective open-heart surgery in clinically proven cases of localized cardiac tamponade (Grumann *et al.* 2012). Our primary goal was to characterize the haemodynamic effects of temporary mechanical compression of the heart and to outline a multi-faceted inflammatory process which may cause potential risks in the post-tamponade phase. This is required to identify the inflammatory mediators, with a potential role in this inflammatory cascade system.

The major finding of Study I is that the post-tamponade period is characterized by a decreased level of systemic perfusion and by an impaired LV function and pulmonary circulation, as evidenced by the MAP, LHEDV, PAP, PVRI and EVLWI data. The pCO<sub>2</sub> gap changes suggested that, in parallel, a significant intestinal microcirculatory dysfunction evolved in this porcine model. More importantly, these responses were associated with abrupt increases in the level of superoxide radical production, big-ET, troponin-T, HMGB1, histamine, intestinal MPO activity and complement activation during the post-tamponade phase.

The cardiac filling disorder induced different vasoconstrictive compensatory reactions in the systemic and pulmonary circulations: the increase in PVRI (112%) was much higher than that in SVRI (32%). Previous experimental and clinical data suggest that the increasing pericardial pressure causes a continuous decline in coronary blood flow due to an increase in coronary vascular resistance (Klopfenstein *et al.* 1987, Skolidis *et al.* 2000). Skolidis *et al.* observed a decreased hyperaemic flow under increased pericardial pressure, which implies an augmented susceptibility to myocardial ischaemia (Skolidis *et al.* 2000). The significantly increased HR during the acute phase of the tamponade could also contribute to the cardiac hypoxia. The decreased CO is compensated by the increased HR, though the length of the systole does not change, while the diastole is shortened. Such a high frequency then results in less time for oxygen delivery to the cardiac muscle cells, and the degree of oxygenation of the heart is therefore worsened. The significantly elevated level of troponin-T during the acute phase of the tamponade also demonstrates the deterioration of oxygenation and damage of the cardiac muscle cells.

After the relief of the tamponade, the MAP was decreased, while the CO was kept compensated, and there were no significant differences as compared with the control group. This may be explained by the normalized preload, as evidenced by the normalized CVP and RHEDV, and the moderate elevation of the afterload. However, the significant decrease in LHEDV indicates a LV dysfunction during the post-tamponade phase. The persistent elevations in PAP and PVRI could contribute to this process, together with the lung oedema, as revealed by the elevated EVLWI. These conclusions are supported by clinical observations on early cardiac failure and pulmonary oedema after removal of the pericardial effusion (Ligero *et al.* 2006, Bernal *et al.* 2007).

In this clinically relevant large animal model, the pericardial tamponade triggered characteristic macro- and microcirculatory changes in the intestines. While the SMA flow, which reflects the blood supply of the small intestine and colon, was diminished during the tamponade, a prolonged impairment of the mucosal microcirculation was detected. This is in accordance with previous assumptions that the macro- and microhaemodynamics may change relatively independently, or may be dissociated in stress conditions (De Backer *et al.* 2010).

In the background of these haemodynamic alterations, a multi-faceted role of humoral mediators, including ET-1, is proposed. Hypoxia is considered to be one of the basic stimuli for ET-1 synthesis. This peptide is produced predominantly by the endothelium, but in pathophysiological states other cell types, such as leukocytes, macrophages, smooth muscle cells, cardiomyocytes and mesangial cells, can also serve as sources of its release (Boros 2003). The increased plasma level of ET-1 could be responsible for the decreased coronary perfusion (Fazekas *et al.* 2001) and pulmonary hypertension. The activation of vasoconstrictor ET receptors can further play a decisive role in acute microcirculatory disorders of the peripheral cardiovascular system. It has been shown that selective ET-A receptor antagonism increases the CO, decreases the peripheral resistance (Wolfard *et al.* 1999, Kaszaki *et al.* 2008) and reduces intestinal microvascular injury and PMN leukocyte accumulation during ischaemia-reperfusion (Wolfard *et al.* 2002).

In addition to its independent role as a dominant vasoconstrictor, the peptide may also influence the functions of other cell types in the circulatory system. ET-1 has been reported to induce leukocyte rolling and adherence through a predominantly ET-A receptor-mediated mechanism (Boros *et al.* 1998). There is a close relationship between a compromised mucosal blood flow and the magnitude of PMN leukocyte-endothelial cell interactions in the intestines (Wolfard *et al.* 2002). On the other hand, ET-1 also causes histamine release from mast cells, which may lead to enhanced vascular permeability and a relative blood loss into the dilated

vessels (Kaszaki *et al.* 2008). Histamine release in the pulmonary circulation contributes to the increase in EVLWI (Walkenstein *et al.* 1985), while in the splanchnic area, histamine release probably plays a role in the counter-regulation of the excessive, prolonged vasoconstriction that contributes to the lethal outcome (Kaszaki *et al.* 1989). Furthermore, ET-1 activates NADPH oxidase, resulting in an increased superoxide radical production (Loomis *et al.* 2005), which can simultaneously reduce NO production, leading to the formation of the highly cytotoxic peroxynitrite (Sheehy *et al.* 1998). From this point of view, the excessive release of ET-1 can be the key player as concerns the spreading inflammatory responses, when intensive complement activation is also ignited. The presence of complement C3 deposits was verified in this tamponade model and we found increased plasma levels of HMGB-1 too, the release of which is additionally directly mediated by the complement cascade. In this scenario, HMGB1 release is a further danger signal to responsive cells, which amplifies the production and secretion of other pro-inflammatory mediators and finally induces excessive inflammation (Yang *et al.* 2002, Ulloa and Messmer 2006).

This study has some limitations. On the one hand, thoracotomy causes severe surgical trauma. A diaphragmatic window through laparotomy could be a possible alternative to reach the pericardial cavity (Park *et al.* 2008). Nevertheless, safe catheterization of the pericardial sac, and quick guidance of the diagnostic instrumentation into the correct positions in the heart cavities are advantages of the open chest model. On the other hand, the inflammatory reaction may be non-specific because cardiogenic and hypovolaemic shock components are not mutually exclusive. Decreases in MAP and CO, splanchnic perfusion and microcirculatory damage may be observed in nearly all forms of circulatory shock (e.g. hypovolaemic, cardiogenic or distributive). Nevertheless, there are also tamponade-specific consequences, such as an elevation of the CVP, a decrease in LHEDV and an increased plasma troponin-T level, which may be direct signs and consequences of tamponade-induced cardiac ischaemia.

In conclusion, we have demonstrated characteristic macrohaemodynamic changes, together with apparent signs of a damaged LV function and a splanchnic inflammatory reaction after the relief of a tamponade. The evidence further suggests that numerous biomarkers, including superoxide radicals, big-ET, HMGB1, histamine, MPO activity or components of the complement system, are significant factors of the inflammatory cascade in this porcine model of pericardial tamponade. On the basis of these results, we assume that these mediators could be promising targets for therapeutic intervention in the future.

## 5.2. Study II

It is widely accepted that inflammatory activation plays a decisive role in low-flow conditions, although the potential of ‘anti-inflammatory’ compounds to prevent or cure low perfusion-induced *in vivo* processes is very limited. With this objective, Study II was designed to develop adjuvant therapy which is able to improve the early haemodynamic and inflammatory changes that occur after restoration of the cardiac tamponade. Numerous experimental results (Ward 2008; Sarma and Ward 2011), together with our findings from Study I, suggested that the components of the complement system could be promising targets for therapeutic intervention. For this purpose, we planned to use the early inhibition of complement C5a with AcPepA, a synthetic, antisense, 17-amino acid peptide acetylated at the N-terminal alanine (Okada *et al.* 2007), during the acute phase of cardiac tamponade. It has been shown that this compound is capable of binding directly to C5a (in its 37-53 amino acid region) in a concentration-dependent manner. The binding between the molecules is very strong (the dissociation requires treatment with 6 M urea *in vitro*) (Fujita *et al.* 2004). AcPepA has proved to be effective in a pilot primate endotoxin shock model (Okada *et al.* 2011).

The major findings of Study II are that the early inhibition of complement C5a with AcPepA effectively restored the impaired LV function and pulmonary circulation in the post-tamponade period. More importantly, these responses were associated with significant decreases in the plasma levels of troponin-T, TNF- $\alpha$ , histamine and complement activation in the tissues after the relief of the cardiac tamponade.

In a relevant study, a significantly increased level of C5a was demonstrated at the beginning of reperfusion, followed by a sudden decrease. The harmful consequences are evoked during this short, but decisive period (Eppinger *et al.* 1997). The relatively short biological half-life (around 30 min) of AcPepA supports the application of this peptide in this time frame. In this line, the study was designed to investigate the effects of a single administration, since its major effects were expected at the very beginning of the reoxygenation. Our IHC examination data clearly revealed that the timing of AcPepA treatment in our pig model was effective, with a decreased level of C3 positivity in the AcPepA-treated tamponade group. These early effects may result in further, longer-term alterations, but as a first step we aimed to identify the key factors in the acute phase of central circulatory failure.

After the relief of the cardiac tamponade, the MAP remained at a decreased level, while the CO did not show any significant differences relative to the control group. This was



due to the elevated HR, which is indicative of the increased strain of the heart muscle. Following AcPepA treatment, the MAP was elevated to the control level and the CO was also maintained. Thus, the most pronounced differences between the compensatory mechanisms of the AcPepA-treated and non-treated tamponade groups were the restored MAP and the lower HR. This seems to be especially crucial if we consider the fact that the circulation of the heart muscle is provided during the diastolic phase, and nearly stops during systole. If the HR is increased, the length of the systole does not change, while the diastole is shortened. The higher the frequency, the shorter the time available for oxygen delivery to the cardiac muscle cells, and therefore a lower HR and maintained CO provide better oxygenation for the heart. The elevated GEDV in the treated group also indicates that the main compensatory mechanism in this early phase is switched from an increased afterload to an elevation of the preload.

It could be supposed that improvements of the LV function and pulmonary circulation play a decisive role in the background of this process. It has been demonstrated that elevation of the PVR reduces the CO, because of the increase in RV afterload leading to an impaired LV function (Avontuur *et al.* 1998). However, a well-timed treatment with AcPepA resulted in decreases in PAP and PVRI, which contributed to the normalizing of LHEDV.

In order to determine the effects of C5a antagonist treatment, we had to consider and rule out artificial influences which may originate from the experimental design. We therefore applied standard fluid replacement therapy in all groups to exclude the effects of the volume status on the macrohaemodynamic parameters. In this line, the favourable haemodynamic changes might be consequences of the reduced biomarker release (TNF- $\alpha$ , histamine and troponin-T) in the post-tamponade phase after AcPepA treatment.

It has been shown that histamine release is induced by a hypoxic condition (Boros 2003) and complement C5a has a direct role in this histamine release from mast cells (El-Lati *et al.* 1994). An elevated histamine level in the plasma may lead to enhanced vascular permeability and a relative blood loss into the dilated vessels, as can occur in the pulmonary circulation in association with the increase in EVLWI (Walkenstein *et al.* 1985). On the other hand, the elevation in plasma histamine level in the early post-tamponade phase shows that this level is higher in the portal venous blood than in the arterial blood (Kaszaki *et al.* 1989). It can decrease the peripheral resistance in patients with circulatory shock (Nagy *et al.* 1986), while in the splanchnic area histamine release probably plays a role in the counter-regulation of the excessive, prolonged vasoconstriction that contributes to the lethal outcome (Kaszaki *et*

*al.* 1989). In the presence of AcPepA, the extent of histamine release was reduced, and this effect too can contribute to the increased venous return.

There is increasing evidence that cytokines in general and TNF- $\alpha$  in particular play an important role in cardiovascular diseases. Thus, increased levels of TNF- $\alpha$  or of its soluble receptors have been implicated in the pathophysiology of ischaemia-reperfusion injury, myocarditis, and the progression of congestive heart failure. TNF- $\alpha$  modulates the two most important haemodynamic determinants of the cardiac function: the peripheral resistance and the cardiac contractility. As concerns the latter, TNF- $\alpha$  could be responsible for a negative inotropic effect, by altering intracellular Ca<sup>2+</sup> homoeostasis and possibly by inducing NO synthesis, which likewise reduces myocyte contractility (Ferrari *et al.* 1999). An increased expression of TNF- $\alpha$  has been demonstrated in a case where infection of the myocardium with Influenza A virus was associated with cardiac tamponade, as a potentially fatal complication (Mamas *et al.* 2007). Our experimental data confirm that an acute cardiac tamponade induces TNF- $\alpha$  release directly, whereas the administration of AcPepA significantly limits its release.

Plasma troponin-T is an indirect, but highly sensitive marker of cardiac tissue ischaemia and has been used to demonstrate the ischaemic effect of a cardiac tamponade (Kelley *et al.* 2009). The lower level of troponin-T following the relief of the cardiac tamponade in the AcPepA-treated tamponade group could be a multiple result of the inhibition of the complement system.

In conclusion, these results demonstrate the relative significance of complement activation in the acute circulatory complications of a cardiac tamponade, and the potential role of C5a antagonism to reduce inflammatory signals which are important components in the development of disturbances of the LV function and the pulmonary circulation.

## **6. SUMMARY OF NEW FINDINGS**

- I. We have used a clinically relevant large animal model to study the haemodynamic and inflammatory changes caused by a cardiac tamponade. A cardiac tamponade induced significant deteriorations of the intestinal macro- and microcirculations, together with impairments of the LV function and pulmonary circulation, associated with potentially harmful inflammatory consequences in the post-tamponade phase.
- II. Definite signs of inflammatory activation were observed with the release of vasoactive and pro-inflammatory mediators, including histamine, big-ET, HMGB-1 and the complement system. Influencing these reactions could possibly offer adjuvant therapeutic targets through which to decrease the complications of cardiogenic shock induced by tamponade.
- III. Via early therapeutic interventions targeting the inhibition of complement C5a with AcPepA, the secondary detrimental circulatory consequences of the cardiac tamponade can be dampened or reversed.

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## 9. ANNEX

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