

Thesis of the PhD dissertation

Implementation of efficient photoautotrophic
H₂ production in the green alga
Chlamydomonas reinhardtii

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Introduction

Since the industrial revolution that began in the 17th century, fossil fuels, including coal, oil and natural gas have been operating the technological and transport networks that drive our society. However, supplying the world with fossil fuels with finite geological reserves threatens our long-term energy supply and puts a huge strain on the environment. Additionally, the energy demand has been increasing worldwide due to population growth. In addition to the problem of providing energy supply, the use of fossil fuels also compromises our own health through chemical and particle pollution. They are not only dangerous to our health, but emissions of carbon dioxide and other greenhouse gases are closely linked to global warming, thus they threaten the long-term sustainability of the Earth's climate as well. Therefore, more attention has to be paid to renewable energy sources.

Natural resources from which energy can be gained and are re-produced within a few years, are called renewable energy sources. Their importance comes from the fact that they do not cause significant pollution and are in line with the principles of sustainable development. Such alternative energy sources include wind energy, geothermal energy, hydro-, biomass- and solar energy. The utilization of renewable energy technologies offers an excellent opportunity to reduce greenhouse gas emissions and slow down the process of global warming by replacing conventional energy sources.

Significant research focuses on the widest possible usage of H₂. H₂ gas is a clean energy carrier, the combustion of which does not produce carbon dioxide, only water. However, about 99% of H₂ is currently produced from fossil fuels, which cause enormous air pollution; therefore, there is a need to develop H₂ production methods based on renewable energy sources. One of the promising ways of doing this is the utilization of green algae, which can produce H₂ with high efficiency linked to their photosynthesis.

Aims of the study

In nature, it has been observed that algae that have been darkened for a few hours produce H₂ for a few minutes upon illumination. The length and efficacy of this short period can be increased by sulfur removal under laboratory conditions, which, however, is accompanied by a number of negative side effects that prevent the industrial application of algal H₂ production.

In the course of my work, I aimed to develop and optimize a technique based on anaerobic induction that is more efficient and less harmful to algal cells. Its elements are as follows:

1. Exploring key steps needed to maintain continuous H₂ production
2. Development and optimization of a novel H₂ production method
3. Testing the sustainability of the system
4. Increasing the efficiency of H₂ production by designing a new type of photobioreactor
5. Identification and characterization of mutant strains involved in photosynthesis with increased H₂ production

Material and methods

1. Maintenance and growth of *Chlamydomonas reinhardtii* strains

In our experiments, we examined and compared the H₂ production of five different strains of *Chlamydomonas reinhardtii*, namely: CC-124, CC-409, L159I-N230Y, *pgr11*, and *pgr5*. Algae were grown in solid or liquid Tris-acetate phosphate (TAP) medium in an algae growth chamber. After three days of cultivation, the cultures were centrifuged and placed in Sueoka's high salt medium (HS). In experiments carried out in 100 ml serum vials, the chlorophyll content was adjusted to 50 µg/ml chl (a+b) and 30 ml of culture were placed in them. In the Roux-type photobioreactor (TCL-PBR) glasses, the chlorophyll content was adjusted to 50 or 150 µg/ml chl (a+b) and 100 ml of culture were placed in the vessels. Before sealing, an O₂ absorbent was placed in special containers in the airspace of the vials. The cultures were placed in the dark for four hours and O₂ and CO₂ were removed from their gas spaces by 20 minutes of N₂ blowing. After four hours of dark anaerobic induction, cultures were placed under 350 or 1000 µmol m⁻² s⁻¹ photon flux density, 23-25 °C and were shaken at a speed of 130 rpm.

2. Chemical treatments

In some experiments, three hours after the cultures were exposed to light, various chemical treatments were applied.

3. O₂ absorbents

The absorbents used in our experiments to remove O₂ from the air were a mixture of sodium chloride and iron. The 504B 100CC Loose (Oxyfree, Tianhua Tech. CO., Ltd. Nanjing, China) mixture was employed in the experiments using serum bottles. The 50CC Loose (O2Zero, Global Reach Ltd, London, UK) absorbent was employed in the experiments using TCL-PBR vials. 1.3 g of O₂ absorbent was added to the serum bottles, while 20 g of O₂ absorbent was used in the TCL-PBR vials.

4. Applied methods:

- Spectroscopic determination of chlorophyll content
- Determination of H₂ and O₂ production by gas chromatography
- Fast chlorophyll *a* fluorescence transient (OJIP) measurements
- Western blot analysis
- Determination of cell number
- *In vitro* hydrogenase activity measurements

Results

1. Exploring key steps needed to maintain continuous H₂ production

To induce hydrogenase expression, a so-called dark anaerobic induction treatment was applied. The *C. reinhardtii* strain CC-124 was chosen as model organism. This strain was already known as an efficient H₂ producer that is capable to produce H₂ under a wide variety of experimental conditions. H₂ production in green algae requires reducing equivalents and anoxic conditions. Reducing equivalents may derived from the photosynthetic electron transport, from starch breakdown and from the subsequent fermentation. The H₂ production has to share the reducing agents pool with competitive reactions. The main competitor for electrons is CO₂ assimilation. The N₂ atmosphere used to ensure anaerobiosis also removes CO₂, the substrate of the Calvin-Benson-Bassham (CBB) cycle, thereby the competitor electron sink is inhibited, allowing increased electron flow towards HydA.

The presence of acetate is necessary for the accumulation of starch, but its metabolism leads to the formation of CO₂, which is able to partially restore the activity of the CBB cycle. To avoid this, we omitted acetate from the medium. In the absence of acetate and so an ample amount of starch, the photosystem II (PSII) -dependent electron transport becomes the main electron source. The water-splitting-based linear electron transport is a more efficient electron source than starch breakdown; however, it is accompanied by O₂ release which prevents the establishment of anoxic conditions. Therefore, to exploit the linear electron transport as a source of sustainable electron source, we opted to remove the released O₂ rapidly and efficiently rather than deactivating PSII by sulfur deprivation. For this purpose, an iron salt-based O₂ absorbent was applied, which reduced O₂ concentration below 0.1% in the headspace of the cultures.

2. Development and optimization of a novel H₂ production method

The method developed by us has fundamental advantages compared to previous methods, namely, under illumination following a few hours of anaerobic dark incubation, H₂ production starts immediately. In contrast to sulfur deprivation, it does not require the replacement of the nutrient solution. It does not depend on the degradation of starch and does not require acetate as it is a photoautotrophic process. Since there is no need for organic carbon sources, the risk of bacterial contamination is much lower. Cultures remain photosynthetically active during H₂ production and can be easily regenerated. The process is based on linear electron transport, the electrons are derived primarily from the water-splitting activity of PSII and has a relatively high light-to-H₂ energy conversion efficiency. In the growth phase, CO₂, which is an industrial by-product, can be used. Relatively high light intensity (320 μmol photon/m²/s) can be used for the production of H₂.

3. Testing the sustainability of the system

The sustainability of our H₂ production system was characterized by the status of the photosynthetic apparatus during a 96-hour experiment. We found that the chl (a+b) content was reduced by only 10% during the experiment. The F_v/F_m value, which is a

parameter of PSII efficacy, showed a slow decrease during the 96-hour period but remained relatively high, above 0.4, both for control samples and in the presence of O₂ absorbent. The amount of PsbA (a major protein of PSII), PetP (a subunit of the cytb₆/f complex), and PsaA (a PSI reaction center protein) remained largely unchanged throughout the experiment in both samples. Since the cells remained photosynthetically active in our experiment, we tried to regenerate them after the H₂ production period and used them again in a new experiment. When these regenerated cells were subjected to a second, dark anaerobic induction period, the yield of H₂ production was similar to that measured in the first cycle.

4. Increasing the efficiency of H₂ production by designing a new type of photobioreactor

We thought that the yield of H₂ production, achieved by anaerobic induction in HS nutrient solution, could be further improved. To achieve this, we used a photobioreactor in which we optimized the gas-liquid as well as the culture volume-surface ratio, the conditions for illumination and algae mixing, and the efficient removal of the produced gases. Thin-Cell-Layer Photobioreactors (TCL-PBRs) are culturing systems with a light path of less than 10 mm and a high surface area-volume ratio. These properties allow TCL-PBR to operate at very high cell densities (1000 µg chl (a+b)/ml culture) and improve photosynthetic efficacy per unit surface of the irradiated area. In our work, to obtain a thin-cell-layer, we kept the alga cultures in a horizontally placed 1-liter Roux bottles (TCL-PBR). The thickness of the cell suspension was approximately 5 mm in the vials. The surface area - volume ratio was increased fourfold (from 0,059 cm⁻¹ to 0,24 cm⁻¹), while the gas-liquid volume ratio was increased threefold (from 3 to 9) relative to serum bottles.

5. Identification and characterization of mutant strains involved in photosynthesis with increased H₂ production

There is great potential in the application of the protocol we have developed for various photosynthetic mutants. After optimization of the photobioreactor, three photosynthetic mutants were tested, which were affected in PSII (L159I-N230Y) or in PSI cyclic electron transport (*pgr11* and *pgr5* mutants). In our experiments sunlight intensity (1000 µmol photon/m²/s) and high chlorophyll content (150 µg/ml) were used.

The H₂ production of all three strains was very similar in the first 24 hours, but the H₂ production of wild-type CC-124 and CC-409 strains showed a continuous decrease, while the L159I-N230Y mutant remained rather stable and it only decreased by day 6. Strain L159I-N230Y produced 30% more H₂ than CC-409, although the condition and performance of their photosynthetic apparatus were similar.

Next, we examined the *pgr5* and *pgr11* mutant strains with the protocol we developed. For the *pgr5* mutant, H₂ production was multiple compared to the wild-type CC-124 strain. The photosynthetic apparatus of the *pgr5* mutant was not significantly damaged even by the intensity of sunlight, and the amount of hydrogenase enzyme was preserved.

Summarizing our results, we can conclude that a new method based on anaerobic induction has been developed, which has several advantages over the previously used sulfur removal protocol. Due to this method and the photobioreactor used in our later experiments, we successfully increased the amount of H₂ production by optimizing the gas-liquid ratio, the cell layer thickness, and the removal of the produced gases. Testing various photosynthetic mutant strains of *Chlamydomonas reinhardtii*, we found that the *pgr5* mutant proved to be the most efficient H₂ producer under the conditions we set up.

Replacing fossil fuels in a way that ensures sustainability and has no environmental or health impact is an increasingly urging problem today. With the obtained knowledge, it may become possible to make use of the H₂-producing capacity of green algae on an industrial scale.

Publications

MTMT identifier: 10055188

Articles which serve the basis for the thesis

Nagy V., Vidal-Meireles A., Podmaniczki A., Szentmihályi K., Rákhely G., Zsigmond L., Kovács L., Tóth S.Z. (2018): The mechanism of photosystem-II inactivation during sulphur deprivation-induced H₂ production in *Chlamydomonas reinhardtii*.

Plant J 94: 548-561.

Impakt faktor: 5,726

Nagy V., Podmaniczki A., Vidal-Meireles A., Tengölics R., Kovács L., Rákhely G., Scoma A., Tóth SZ. (2018): Water-splitting-based, sustainable and efficient H₂ production in green alga as achieved by substrate limitation of the Calvin-Benson-Bassham cycle. Biotechnol Biofuels 11: 69

Impakt faktor: 5,452

Nagy V.* Podmaniczki A.*, Vidal-Meireles A., Kuntam S., Herman É., Kovács L., Tóth D., Scoma A., Tóth SZ. (2021): Thin cell layer cultures of *Chlamydomonas reinhardtii* *L159I-N230Y*, *pgr11* and *pgr5* mutants perform enhanced hydrogen production at sunlight intensity. Bioresour Technol 333: 125217

Impakt faktor: 11,889

Podmaniczki A., Nagy V., Vidal-Meireles A., Tóth D., Patai R., Kovács L., Tóth SZ. (2021): Ascorbate inactivates the oxygen-evolving complex in prolonged darkness. Physiol Plantarum 171: 232-245

Impakt faktor: 5,081

Széles E., Nagy K., Ábrahám Á., Kovács S., Podmaniczki A., Nagy V., Kovács László., Galajda P., Tóth SZ. (2022): Microfluidic platforms designed for morphological and photosynthetic investigations of *Chlamydomonas reinhardtii* on a single-cell level

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