

DEPARTMENT OF PEDIATRICS
ALBERT SZENT-GYÖRGYI MEDICAL UNIVERSITY
SZEGED, HUNGARY

THESIS

EFFECT OF ALLOPURINOL TREATMENT OF PREECLAMPTIC
MOTHERS ON THE ANTIOXIDANT CAPACITY OF THE NEWBORN
(A HYPOTHESIS AND METHODOLOGICAL APPROACH)

ESZTER PAPP



1992

B 518



P 2149



CONTENTS

ABBREVIATIONS	1
INTRODUCTION	1
PATIENTS AND METHODS	6
RESULTS	14
DISCUSSION	14
ACKNOWLEDGEMENTS	18
REFERENCES	19

EFFECT OF ALLOPURINOL TREATMENT OF PREECLAMPTIC MOTHERS
ON THE ANTIOXIDANT CAPACITY OF THE NEWBORN
(A HYPOTHESIS AND METHODOLOGICAL APPROACH)

ABBREVIATIONS

CPO, cumene hydroperoxide

EDRF, endothelium derived relaxing factor

GSH, reduced form of glutathione

GSSG, oxidized form of glutathione

NTBI, non-transferrin bound iron

OH[·], hydroxyl radical

PG I₂, prostacyclin

TRAP, total free radical trapping antioxidant parameter

TX A₂, thromboxane A₂

INTRODUCTION

Preeclampsia, a frequent disorder of human pregnancy, characterized by persistent arterial hypertension and proteinuria appearing after the 20th week of gestation, is one of the most important causes of premature birth. Although the syndrome is common and well known, its pathogenesis is still unclear. So far there have been two major hypotheses offered to explain the generalized vasoconstriction found in preeclampsia (Fig.1).

The first one suggests that there is an imbalance in the production of vasoactive prostaglandines caused by circulating lipid peroxidation products (Fig.2). Prostacyclin (PG I₂) is a vasodilator and inhibitor of platelet aggregation, whereas thromboxane A₂ (TX A₂) is a vasoconstrictor and a promoter of platelet aggregation. Circulating products of the excessive lipid peroxidation in preeclampsia inhibit the synthesis of prostacyclin via inhibition of prostacyclin synthetase enzyme. Thromboxane A₂ synthesis, however, is not negatively

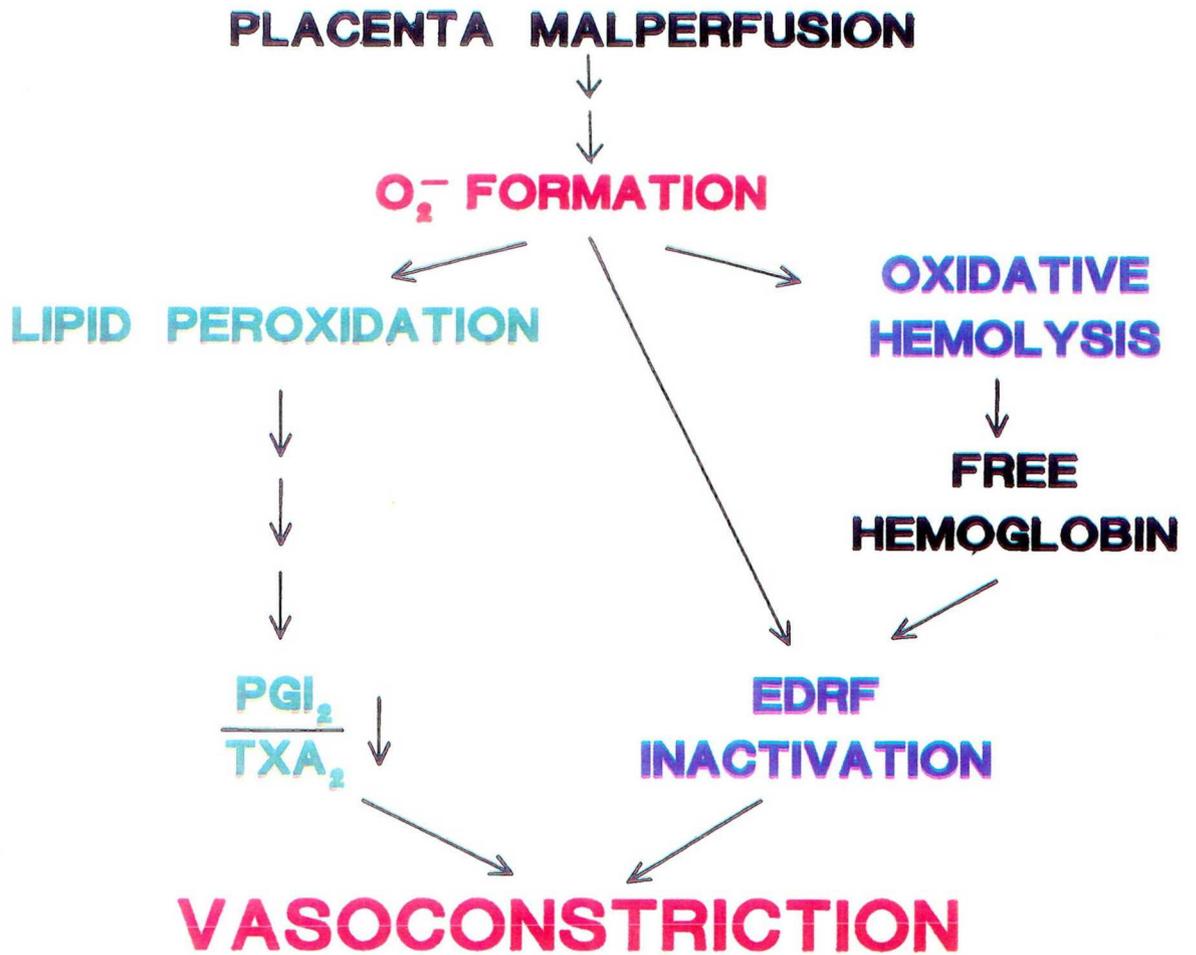


Figure 1.

Hypotheses to the pathomechanism of preeclampsia

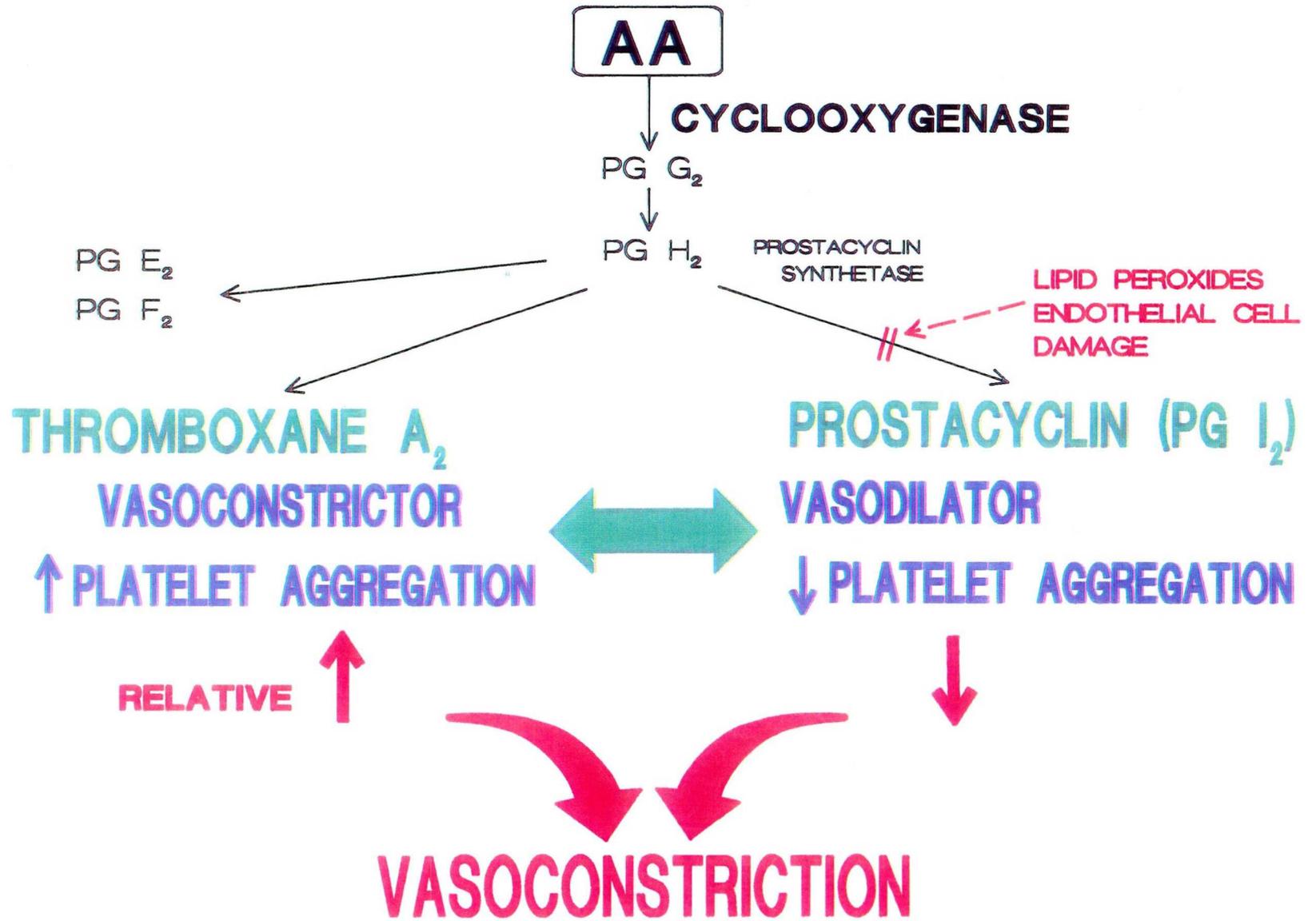


Figure 2. The role of vasoactive prostaglandines in the pathomechanism of preeclampsia

influenced by lipid peroxides. This results in an imbalance between the vasoconstrictor TX A₂ and vasodilator PG I₂, in which the former dominates resulting in a generalized vasoconstriction (2,29,30).

According to the other hypothesis free hemoglobin as well as superoxide free radicals inactivate the endothelium derived relaxing factor (EDRF) produced by vascular endothelial cells (2,3,31). EDRF, identified as either nitric oxide or a related compound, plays an important role in the control of tissue blood flow. In preeclampsia the increased amount of free hemoglobin (coming probably from the increased rate of oxidative hemolysis) binds to, and inhibits the effects of EDRF, which results in vasoconstriction (Fig 1.).

We offer the hypothesis that in the background of both mechanisms there may be a free radical mediated oxidative stress.

The primary defect in preeclampsia has been shown to be a severely depressed uteroplacental blood flow (1) with a consecutive tissue hypoxia. It is well known that in hypoxic tissues hypoxanthine is accumulated as a result of the increased ATP breakdown. At the same time xanthine dehydrogenase enzyme, normally present in endothelial cells, is transformed into xanthine oxidase (Fig 3.). In the presence of this enzyme superoxide free radicals are formed. These products can be toxic in themselves but their toxicity increases greatly in the presence of iron, as iron catalyzes the formation of the extremely reactive hydroxyl radicals (OH·) in the iron catalyzed Haber-Weiss reaction:



The severity of the damage caused by free radicals depends on the actual antioxidant status of the body. One of the most important superoxide scavengers in erythrocytes is the glutathione redox system. During oxidative stress GSH, the reduced form, is oxidized and GSSG is formed, therefore there is a rapid decrease in the amount of GSH (Fig.3.). GSSG is then reduced by the NADPH-dependent glutathione reductase enzyme. Thus, it is not only the total amount of GSH that is important in the antioxidant defence system, but also the capacity of erythrocytes to regenerate GSH after an oxidative stress.

THE XANTHINE OXIDASE ENZYME SYSTEM IN HYPOXIC TISSUES

ANTI OXIDANTS

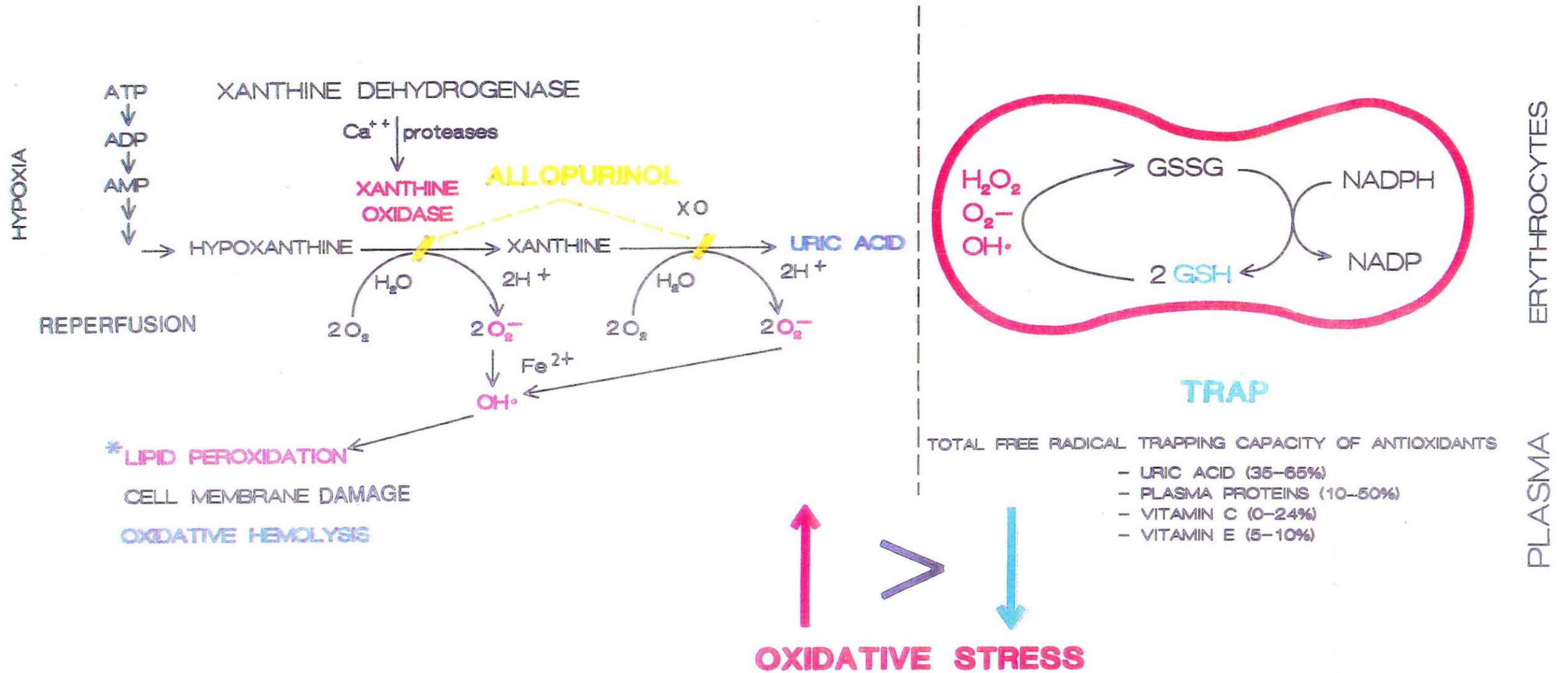


Figure 3.

There are several antioxidants in the plasma as well, e.g. uric acid, SH groups of plasma proteins, vitamin E and vitamin C. TRAP, the total free radical trapping capacity of plasma antioxidants is the result of the synergistic action of these antioxidants (4,5).

Although a variety of the above mentioned antioxidants serve to control the harmful effect of free radicals, under certain conditions the protective mechanisms can be overwhelmed. This imbalance between oxidant and antioxidant forces in which the former dominates is defined as oxidative stress, resulting in uncontrolled lipid peroxidation and cell membrane damage (Fig.3.).

Our hypothesis about the possible pathogenic role of an increased xanthine oxidase enzyme activity generating superoxide free radicals seems to be supported by the fact that in many preeclamptic pregnancies not only hyperuricaemia, but also an elevated serum level of lipid peroxidation products and an increased rate of hemolysis has been found which can all indicate an oxidative stress.

If, indeed, a free radical induced oxidative stress is in the background of preeclampsia, then allopurinol, a specific xanthine oxidase enzyme inhibitor must have a beneficial effect on the newborn. By reducing superoxide generation allopurinol decreases the oxidative stress on the infant.

As in Holland allopurinol is given as an adjuvant treatment to some preeclamptic mothers with hyperuricaemia, it has given us the chance to evaluate its effect on the newborn.

PATIENTS AND METHODS

Our patients fall into 3 groups (Fig.4). All of them are prematures, their gestational ages are between 30 and 35 weeks. In the first 2 groups there are prematures born from preeclamptic pregnancies, mothers of babies in group II received allopurinol treatment during pregnancy. (The usual dosage of allopurinol: if serum uric acid level is above 0.36 mmol/l then 300-900 mg daily.) Babies in the control group are born from normal pregnancies, but prematures.

Venous cord blood samples are taken from the separated placenta within 15 minutes after birth. The blood is collected over Na₂EDTA and immediately centrifuged (800xg for 10 min). The supernatant (plasma) is stored at -70°C for later determination of TRAP and uric acid concentrations (d),e), see below), whereas measurements from the packed cells (a),b),c), see below) are carried out immediately.

In order to obtain information about the actual antioxidant status of our patients we have planned to determine the following parameters (Fig.5.):

- a) total GSH concentration of erythrocytes indicating the 'in vivo' oxidative stress;
- b) the capacity of erythrocytes to regenerate GSH after an 'in vitro' oxidative stress;
- c) the proportion of hemolysis after an 'in vitro' oxidative stress as an indicator of the oxidative susceptibility of erythrocytes;
- d) TRAP, as a parameter of the total free radical trapping capacity of plasma antioxidants;
- e) serum uric acid concentrations indicating the xanthine oxidase enzyme activity.

The total GSH concentration of erythrocytes and the capacity of erythrocytes to regenerate GSH after an 'in vitro' peroxidative stress are both determined by the glutathione stability test according to Koster et al (7) (Fig.6.). Separated packed cells are washed 3 times with a 0.1 M phosphate buffer containing 10 mM glucose (pH=7.4) and diluted with the same buffer solution. The cells are then exposed to cumene hydroperoxide (CPO) at concentrations of 0.75 mM and 1.5 mM. They are incubated at 37°C in shaking waterbath for 60 minutes. Parallel control studies are performed by means of incubation of cells in the buffer solution alone. Samples are taken at 0, 5, 10, 15, 20, 25, 30, 40, 50, 60 minutes and reduced glutathione (GSH) concentrations are determined from them with the GSH assay according to Beutler et al (39) based on the reduction of dithiobis-nitrobenzoic acid (DTNB). GSH concentrations at the indicated time points are calculated and expressed as a percentage of the total GSH concentration of erythrocytes.

During the control studies GSH concentrations remained constant, giving the original total GSH concentrations of erythrocytes, expressed in nmol/ml packed cells.

METHODS

ERYTHROCYTES

PLASMA

CUMENE HYDROPEROXIDE

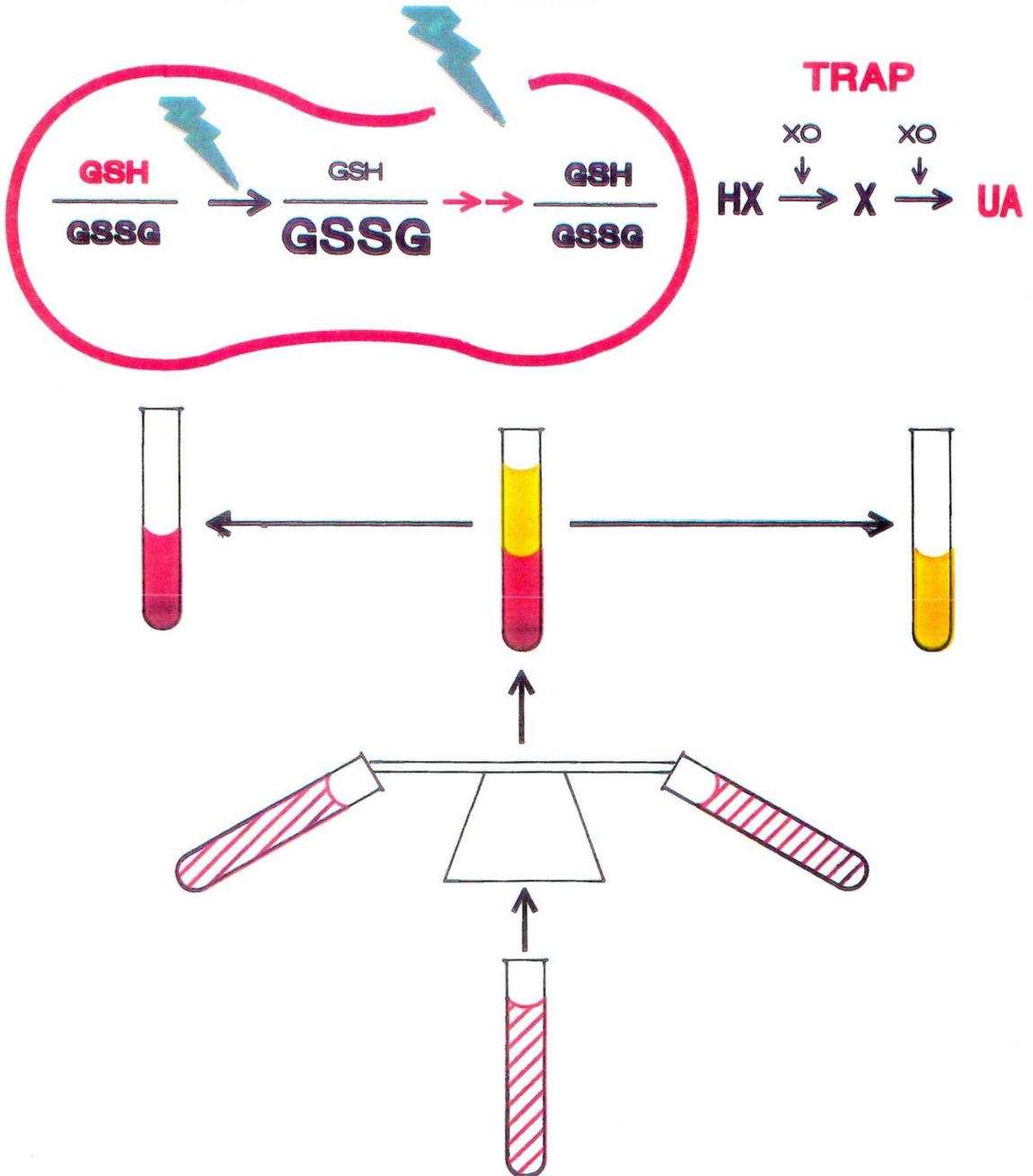


Figure 5.

GSH ASSAY (Koster et al.)

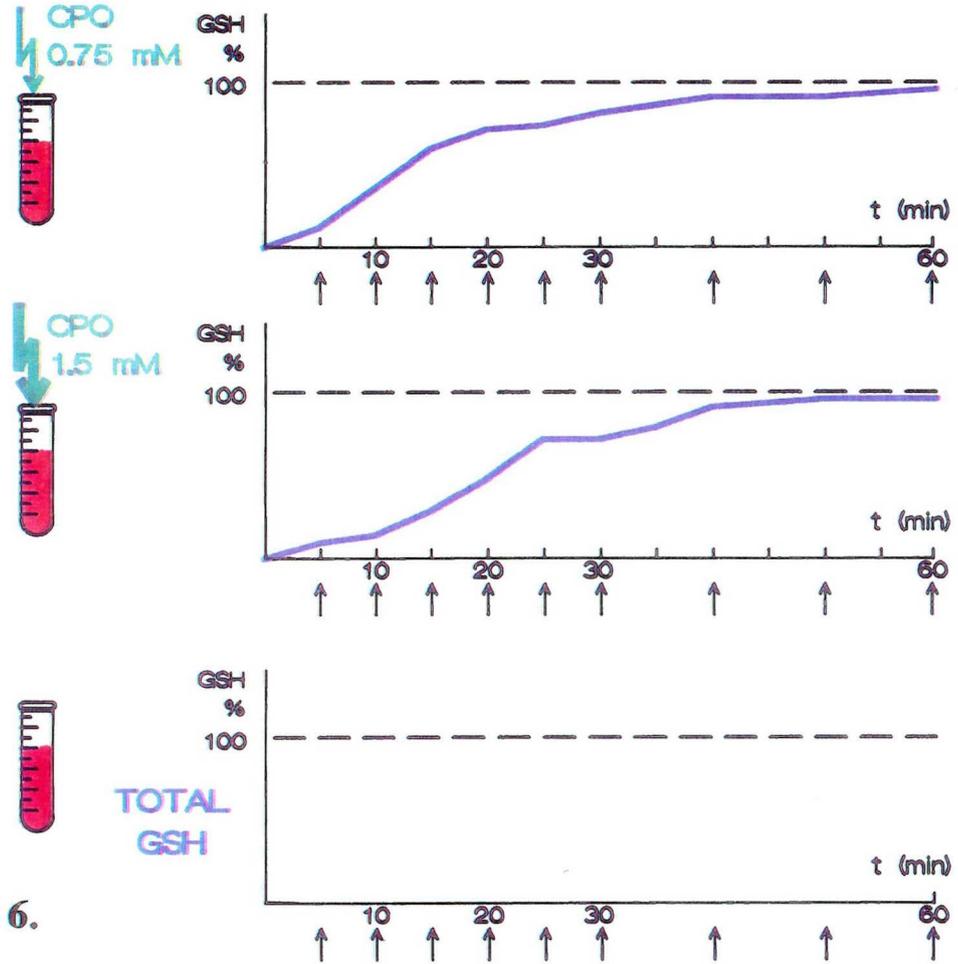


Figure 6.

The oxidative susceptibility of erythrocytes, indicated by the rate of hemolysis after an 'in vitro' peroxidative stress is determined as follows (Fig.7.): Washed, diluted packed cells are given a peroxidative stress with CPO (1.5 mM). After 60 minutes' incubation at 37°C the sample is centrifuged and hemoglobin concentration of the supernatant is determined with a hemoglobin reagent (containing 100 mg NaCN + 300 mg K₃Fe(CN)₆/L 10 mM phosphate buffer, pH=7.4), absorptions are read at 546 nm. Total hemoglobin concentration of the packed cells is determined with the same method after they are hemolyzed in a hypotonic solution. The rate of hemolysis due to the 'in vitro' peroxidative stress is calculated by comparing the supernatant's hemoglobin concentration to the total hemoglobin concentration of the packed cells.

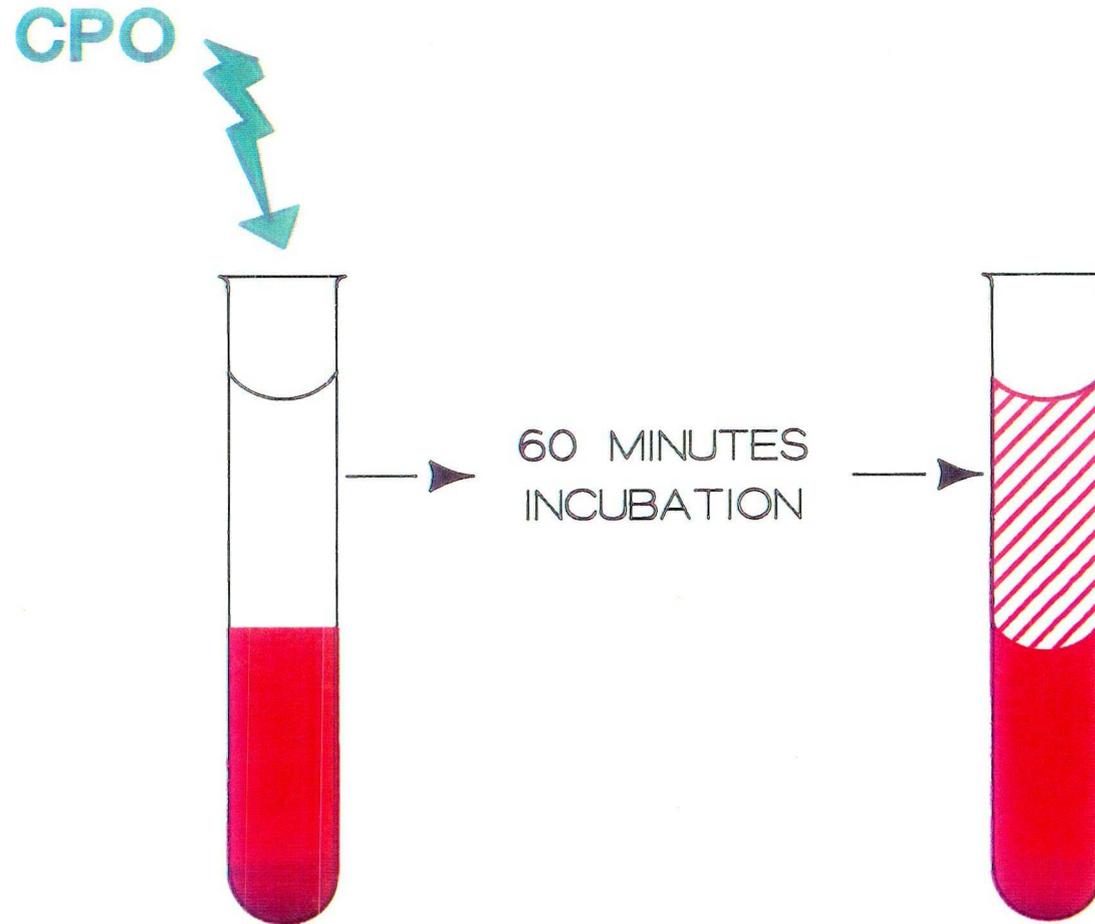
TRAP, as a parameter of the total free radical trapping capacity of plasma antioxidants was measured with the TRAP assay (4,5,6,8). The principle of the measurement is as follows (Fig.8.):

A buffer solution contains linoleic acid, as the substrate of controlled lipid peroxidation, ABAP (2,2'-azo-bis(amidopropane)), a thermal labile free radical initiator and the plasma sample containing a certain amount of antioxidants. The system is closed, oxygen concentration is continuously measured with a Clarke-type oxygen electrode.

Without the antioxidants free radicals would rapidly be consumed for lipid peroxidation, and thus the total oxygen concentration of the system would fall steeply. But as long as the added plasma has a high trapping capacity lipid peroxidation is inhibited - therefore the fall in the total oxygen concentration of the system is not so steep. The higher its antioxidant capacity the longer the plasma can inhibit lipid peroxidation. Antioxidant activity of the serum is therefore measured by the time taken to prevent maximum oxygen uptake. The system is calibrated by the addition of a known amount of water soluble vitamin E analogue (Trolox=6 hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid) with a known trapping capacity.

The serum uric acid concentration is determined as an indicator of the xanthine oxidase enzyme activity.

OXIDATIVE SUSCEPTIBILITY OF ERYTHROCYTES



$$\% \text{ OF HEMOLYSIS} = \frac{\text{HAEMOGLOBIN CONCENTRATION IN SUPERNATANT}}{\text{TOTAL HAEMOGLOBIN CONCENTRATION}} \times 100$$

Figure 7.

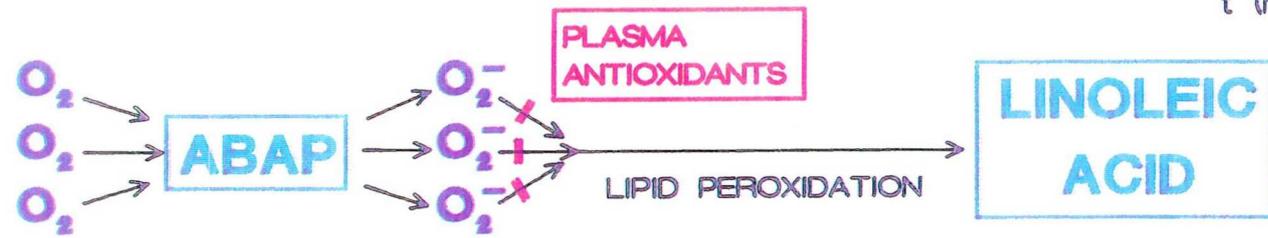
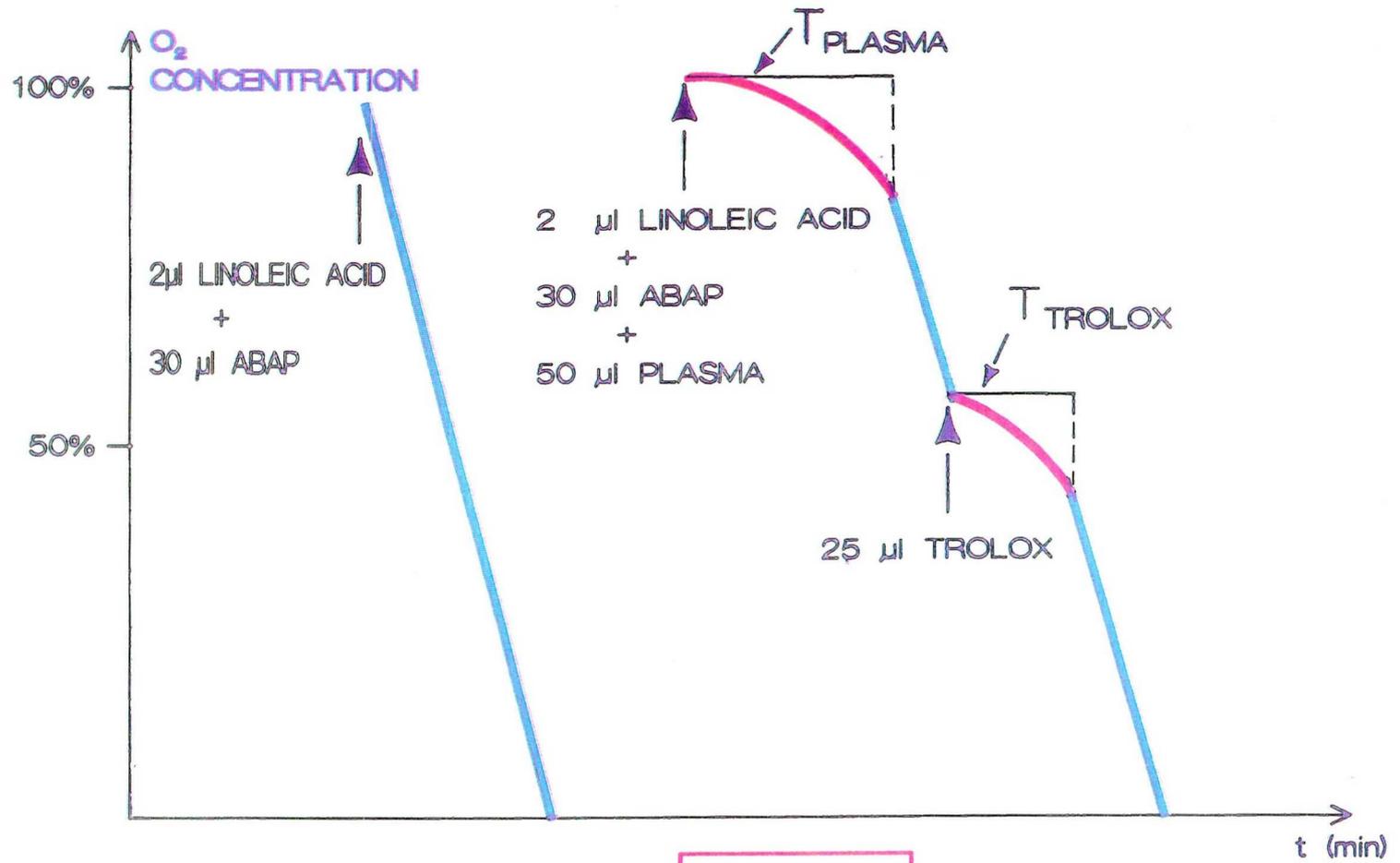


Figure 8.

RESULTS

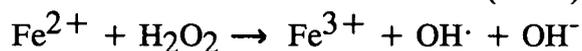
The aim of this study is to introduce the idea of the possible beneficial effect of allopurinol treatment in preeclampsia on the basis of various publications, and to present the methods, already elaborated, which will enable us to find proof to this hypothesis. However, some initial results obtained with methods a)-c) are shown in Table 1.

In all 3 patient groups the initial GSH concentration of the erythrocytes was restored within 60 minutes after the 'in vitro' peroxidative stress, no matter which concentration of CPO was used. There tends to be some difference, however, in the erythrocytes' GSH concentrations indicating a stronger 'in vivo' oxidative stress in the non-treated preeclamptic patient group, which seems to have been prevented by allopurinol treatment. The rate of hemolysis after the 'in vitro' peroxidative stress shows some difference as well in patient groups I and II, suggesting a higher fragility, a higher oxidative susceptibility of erythrocytes in the non-treated patient group.

DISCUSSION

In the present study it is suggested that adjuvant allopurinol treatment of preeclamptic mothers may result in a decrease in the oxidative stress on the baby, a decrease in the oxidative susceptibility of its erythrocytes and, consequently, an improvement of the newborn's clinical condition.

It has been suggested by several authors that the most severe oxidative damage is caused not only directly by superoxide anions, but also by the even more reactive hydroxyl radicals ($\text{OH}\cdot$) (10,11,12,13,14,15,17,18,19). They are generated in the presence of non-transferrin-bound iron (NTBI) (9) in the Fenton reaction:



Iron that is not bound to the usual transport protein but to different extracellular low molecular weight compounds is referred to as NTBI. It becomes elevated when transferrin saturation is high and transferrin is no longer able to bind iron. (Excessive parenteral iron administration during pregnancy may also be

RESULTS

Table 1.

I. P R E E C L A M P S I A

II.

III.

C O N T R O L

ALLOPURINOL



TOTAL GSH CONCENTRATION	<p>1875.0 nmol/ml packed cells</p> <p>2543.8 nmol/ml packed cells</p>	<p>3089.7 nmol/ml packed cells</p> <p>3223.2 nmol/ml packed cells</p>	<p>3308.1 nmol/ml packed cells</p> <p>3467.4 nmol/ml packed cells</p> <p>2629.9 nmol/ml packed cells</p>
% OF HEMOLYSIS	<p>22.8 %</p> <p>71.0 %</p>	<p>1.9 %</p> <p>5.9 %</p>	<p>0.4 %</p> <p>9.1 %</p> <p>9.6 %</p>
GSH STABILITY			

considered as a possible factor leading to iron overload.)

Considering the fact that superoxide (generated by the xanthine oxidase enzyme system) and other free radicals can release iron from ferritin (10,15,16,18), it is possible that the oxidative damage in the newborn is, at least partly, caused by hydroxyl radicals.

It has also been proved that the erythrocytes of the newborn are more susceptible to oxidative stress and there is a high tendency of hemoglobin oxidation in neonates (20,21,22). The oxidative derivatives of hemoglobin: oxihemoglobin and methemoglobin are able to generate $\text{OH}\cdot$ and another, also highly damaging 'reactive species'(13) - they may account for the most severe oxidative damage in neonates.

By reducing the superoxide radical generation with allopurinol treatment we might reduce the iron release from ferritin and the production of oxidized hemoglobin derivatives, thus preventing the formation of the highly reactive $\text{OH}\cdot$ radicals.

Allopurinol treatment has already been found to be beneficial both in human (26,38) and in experimental animal (27) subjects endangered by hypoxia and oxidative stress. What we suggest in this study is practically an allopurinol treatment started before birth. The question must be raised whether allopurinol passes through the placental barrier and can in fact have a therapeutic effect on the baby 'in utero'. No data concerning this particular problem have been found in literature. A relatively simple micromethod, however, suitable for measuring blood levels of allopurinol and its xanthine oxidase inhibitory activity has been developed and published (28) and with this method allopurinol was detected in the blood of infants following allopurinol treatment during pregnancy (Boda,D., unpublished data).

In spite of the fact that xanthine oxidase enzyme activity can be determined directly with the caffeine test (25), we suggest serum uric acid determination as an indicator of xanthine oxidase enzyme activity. Considering the fact that we use cord blood for our measurements, it is obvious that the caffeine test cannot be performed. On the other hand, serum uric acid levels have been shown to correlate well with the results of the caffeine test (25), therefore we consider serum

uric acid level to be a parameter sufficiently informative on xanthine oxidase enzyme activity.

Many investigators reported increased serum uric acid levels in women with preeclampsia (33), and also a positive correlation was demonstrated between the magnitude of serum uric acid levels, clinical severity of preeclampsia and perinatal outcome (34-37). It was also found that serum uric acid levels may begin to rise before the appearance of hypertension and proteinuria (32), meaning that the rise in the serum level of uric acid is probably not caused by renal impairment but by the increased xanthine oxidase activity.

Although in the present study GSH concentration of erythrocytes is used as an indicator of the 'in vivo' oxidative stress, it has been suggested (23,24) that the ratio of oxidized/reduced glutathione (GSSG/GSH) is a much more sensitive and reliable indicator. However, the determination of the latter is much more complicated, therefore it was purely for technical reasons that GSH determination was chosen.

The defences against oxygen toxicity include enzymes (e.g. superoxide dismutase, glutathione peroxidase), metal chelators (transferrin, ceruloplasmin) and various plasma antioxidants e.g. vitamin E (VE), vitamin C (VC), sulfhydryl groups (SH), uric acid (UA), etc. (5). Synergistic interactions occur between these various antioxidants and therefore complete understanding of the total antioxidant defence system in the baby requires measurement of the combined effect of the antioxidants and not just the measurement of their individual concentrations (4). There are two major types of antioxidants. Those that reduce the rate of production of new radicals, e.g. transferrin, allopurinol, are referred to as primary or preventive antioxidants. Antioxidants that trap radicals and thereby reduce the chain length of oxidation are referred to as secondary or chain-breaking antioxidants, e.g. vitamin E and C (5). The TRAP provides information on the total capacity of the chain-breaking (secondary) antioxidants in the plasma. TRAP can either be measured directly with the TRAP assay ($TRAP_{meas}$), or calculated from the measured concentrations of the individual plasma antioxidants by using their experimentally determined efficiency values (8) as follows:

$$\text{TRAP}_{\text{calc}}(\mu\text{mol/l}) = 1.3[\text{UA}] + 1.7[\text{VC}] + 2.0[\text{VE}] + 0.2[\text{SH}]$$

It has been found however, that the values of $\text{TRAP}_{\text{calc}}$ were considerably lower than the $\text{TRAP}_{\text{meas}}$ (5). The reason for this can be on one hand the fact, that not all of the plasma antioxidants have been identified yet. On the other hand, the $\text{TRAP}_{\text{calc}}$ does not take into account the very important interactions that occur between antioxidants when they work in unision (4). For example, the recycling of vitamin E by vitamin C is believed to be an important mechanism in defending against lipid peroxidation (4). These interactions do occur when the $\text{TRAP}_{\text{meas}}$ assay is performed and may explain the greater than predicted free radical trapping capacity in plasma (5). Determination of $\text{TRAP}_{\text{meas}}$ therefore gives reliable information on the actual antioxidant capacity of the plasma taking all (even the unidentified) antioxidants as well as their synergistic actions into account.

Our initial results, as well as data from publications suggest that allopurinol treatment of preeclamptic mothers may have a beneficial effect on their babies probably due to the specific inhibition of xanthine oxidase enzyme and thus the reduced generation of reactive superoxide radicals. Further investigations in order to find out more about the pathogenesis of preeclampsia and the possible role of oxygen toxicity in it are in progress.

ACKNOWLEDGEMENTS

The author would like to thank **Dr. Wim Sluiter** (Dept. of Biochemistry I., Erasmus University, Rotterdam, The Netherlands), **Dr. Iona Németh** (Dept. of Pediatrics, Albert Szent-Györgyi Medical University, Szeged, Hungary) and **Dr. John N. van den Anker** (Sophia Children's Hospital, Erasmus University, Rotterdam, The Netherlands) for their help and advice.

This work was supported in part by the TEMPUS program.

REFERENCES

1. Losonczy,G., Todd,H., Palmer,D.C., Hertelendy,F. Prostaglandins, norepinephrine, angiotensin II and blood pressure changes induced by uteroplacental ischemia in rabbits. *Clin. exp. Hypert.-Hypert. in Pregnancy*, **B** 5, 271, 1986-87.
2. Hubel,C.A., Roberts,J.M., Taylor, R.N., Musci,T.J., Rogers,G.M., McLaughlin,M.K. Lipid peroxidation in pregnancy. *Am. J. Obstet. Gynecol.*, 161, 1025, 1989.
3. Sarrel,P.M., Lindsay, D.C., Poole-Wilson,P.A., Collins,P. Hypothesis: inhibition of endothelium-derived relaxing factor by haemoglobin in the pathogenesis of preeclampsia. *Lancet*, 336, 1030, 1990.
4. Wayner,D.D.M., Burton,G.W., Ingold,K.U., Barclay,L.R.C., Locke,S.J. The relative contributions of vitamin E, urate, ascorbate and proteins to the total peroxy radical-trapping antioxidant activity of human blood plasma. *Biochim. Biophys. Acta*, 924, 408, 1987.
5. Lindeman,J.H.N., Zoeren-Grobben,D., Schrijver,J., Speek,A.J., Poorthuis, B.J.H.M., Berger,H.M. The total free radical trapping ability of cord blood plasma in preterm and term babies. *Pediatr.Res.*, 26, 20, 1989.
6. Wayner,D.D.M., Burton,G.W., Ingold,K.U., Locke,S. Quantitative measurement of the total, peroxy radical-trapping antioxidant capability of human blood plasma by controlled peroxidation. *FEBS Letters*, 187, 33, 1985.
7. Koster,J.F., Biemond,P., Swaak,A.J.G. Intracellular and extracellular sulphhydryl levels in rheumatoid arthritis. *Ann. Rheum.Dis.*, 45, 44, 1986.
8. Thurnham,D.I., Situnayake,R.D., Kootatthep,S., McConkey,B., Davies,M. Antioxidant status measured by the TRAP assay in rheumatoid arthritis. In: Rice Evans C (ed) *Free Radicals, Oxidant Stress and Drug Action*. Richelieu Press, London, pp.161-185, 1987.
9. Wang,W.C., Ahmed,N., Hanna,M. Non-transferrin bound iron in long-term transfusion in children with congenital anemias. *J. Pediatr.*, 108, 552, 1986.
10. Monteiro,H.P., Winterbourn,C.C. The superoxide-dependent transfer of iron from ferritin to transferrin and lactoferrin. *Biochem. J.*, 256, 923, 1988.

11. Berger,H.M., Lindeman,J.H.N., vanZoeren-Grobbe,D., Houdkamp,E., Schrijver,J., Kanhai,H.H. Iron overload, free radical damage, and rhesus hemolytic disease. *Lancet*, 335, 933, 1990.
12. Halliwell,B. Protection against tissue damage in vivo by desferroxamine: what is its mechanism of action? *Free Radical Biol. Med.*, 7, 645, 1989.
13. Puppo,A., Halliwell,B. Formation of hydroxyl radicals from hydrogen peroxide in the presence of iron. *Biochem. J.*, 249, 185, 1988.
14. Biemond,P., Swaak,A.J.G., van Eijk,H.G., Koster,J.F. Intraarticular ferritin-bound iron in rheumatoid arthritis. *Arthritis Rheum.*, 29, 1187, 1986.
15. Biemond,P., Swaak,A.J.G., van Eijk, H.G., Koster,J.F. Superoxide dependent iron release from ferritin in inflammatory diseases. *Free Radical Biol. Med.*, 4, 185, 1988.
16. Biemond, P., van Eijk, H.G., Swaak, H.J.G., Koster, J.F. Iron mobilization from ferritin by superoxide derived from stimulated polymorphonuclear leukocytes: possible mechanism in inflammation diseases. *J. Clin. Invest.*, 73, 1576, 1984.
17. Biemond,P., Swaak,A.J.G., van Eijk,H.G., Koster,J,F. Intraarticular ferritin-bound iron in rheumatoid arthritis. *Arthritis Rheum.*, 29, 1187, 1986.
18. Biemond,P., Swaak,A.J.G., Beindorff,C.M., Koster,J.F. Superoxide-dependent and -independent mechanisms of iron mobilization from ferritin by xanthine oxidase. *Biochem. J.*, 239, 169, 1986.
19. Halliwell,B., Gutteridge,M.C. Role of free radicals and catalytic metal ions in human disease: an overview. *Meth. Enzymol.*, 186, 1, 1990.
20. Shahal,Y., Bauminger,E.R., Zmora,E., Katz,M., Mazor,D., Horn,S., Meyerstein,N. Oxidative stress in newborn erythrocytes. *Pediatr. Res.*, 29, 119, 1991.
21. Etukudo,M.H., Ramachandran,M., Iyer,G.Y.N. Methemoglobin formation and glutathione disappearance in cord blood red cells exposed to acetylphenylhydrazine. *Clin. Chim. Acta*, 138, 135, 1984.
22. Jain,S.K. The neonatal erythrocyte and its oxidative susceptibility. *Semin. Hematol.*, 26, 286, 1989.
23. Németh,I., Boda,D. The ratio of oxidized/reduced glutathione as an index of

- oxidative stress in various experimental models of shock syndrome. In: Matkovics,B., Karmazsin,L., Kalasz,H. (eds) Radicals, ions and tissue damage. Akadémiai Kiadó, Budapest, p.41, 1990.
24. Németh,I., Boda,D. Oxidized and reduced glutathione levels in blood samples from premature infants with RDS and critically ill children. In: Matkovics,B., Boda,D., Kalász,H. (eds) Oxigen free radicals and the tissue injury. Akadémiai Kiadó, Budapest, p.269, 1988.
 25. Boda,D., Németh,I. Measurement of urinary caffeine metabolites reflecting the 'in vivo' xanthine oxidase activity in premature infants with RDS and in hypoxic states of children. In: Matkovics,B., Karmazsin,L., Kalász,H. (eds) Radicals, ions and tissue damage. Akadémiai Kiadó, Budapest, p.35, 1990.
 26. Boda,D., Németh,I., Hencz,P., Dénes,K. Effect of allopurinol treatment in premature infants with idiopathic respiratory distress syndrome. *Dev. Pharmacol. Ther.* 7, 357, 1984.
 27. Jenkinson,S.G., Roberts,R.J., DeLemos,R.A., Lawrence,R.A., Coalson,J.J., King,R.J., Null,D.M., Gerstmann,D.R. Allopurinol-induced effects in premature baboons with respiratory distress syndrome. *J. Appl. Physiol.*, 70, 1160, 1991.
 28. Németh,I., Boda,D. A method for determination of the inhibition of xanthine oxidase activity in plasma during allopurinol treatment. *Europ. J. Clin. Pharmacol.*, 13, 423, 1978.
 29. Schiff,E., Barkai,G., Mashiach,S. Arachidonic acid metabolism in the pathophysiology and prevention of preeclampsia - a review. *Isr. J. Med. Sci.*, 27, 578, 1991.
 30. Schiff,E., Peleg,E., Goldenberg,M., Rosenthal,T., Ruppin,E., Tamarkin,N., Barkai,G., Ben-Baruch,G., Yahal,I., Blankstein,J., Goldman,B., Mashiach,S. The use of aspirin to prevent pregnancy-induced hypertension and lower the ratio of thromboxane A₂ to prostacyclin in relatively high risk pregnancies. *N. Engl. J. Med.*, 321, 351, 1989.
 31. Edwards,D.H., Griffith,T.M., Ryley,H.C., Henderson,A.H. Haptoglobin-haemoglobin complex in human plasma inhibits endothelium dependent relaxation: evidence that endothelium derived relaxing factor acts as a local

- autocoid. ardiovasc. Res., 20, 549, 1986.
32. Dekker,G.A., Sibai,B.M. Early detection of preeclampsia. Am. J. Obstet. Gynecol., 165, 160, 1991.
 33. Stander,H.J., Duncan,E.E., Sisson,W.E. Chemical studies in toxaeimias of pregnancy. Bull. John Hopkins Hosp., 36, 411, 1925.
 34. Lancet,M., Fisher,I.L. The value of blood uric acid levels in toxaeimia of pregnancy. J. Obstet. Gynecol. Br. Emp., 63, 116, 1956.
 35. Liedholm,H., Montan,S., Aberg,A. Risk grouping of 113 patients with hypertensive disorders during pregnancy, with respect to serum urate, proteinuria, and time of onset of hypertension. Acta Obstet. Gynecol. Scand., 118(suppl), 43, 1984.
 36. Redman,C.W.G., Beilin,L.J., Bonnar,J., Wilkinson,R.H. Plasma-urate measurement in predicting fetal death in hypertensive pregnancy. Lancet, 1, 1370, 1976.
 37. Schuster,E., Wepelmann,B. Plasma urate measurements and fetal outcome in preeclampsia. Gynecol. Obstet. Invest., 12, 162, 1981.
 38. Boda,D., Németh,I. Effect of parenteral allopurinol teratment of critically ill children in need of intensive care. Acta Paed. Hung., 24, 247, 1983.
 39. Beutler,E., Duron, O., Kelly,B.M. Improved method for the determination of blood glutathione. J. Lab. Clin. Med., 61, 882, 1963.
 40. Halliwell,B. Tell me about free radicals, doctor: a review. J. Royal Soc. Med., 82, 747, 1989.
 41. Saugstad, O.D. Oxygen toxicity in the neonatal period. Acta Paed. Scand., 79, 881, 1990.
 42. Sullivan, J.L. Iron, plasma antioxidants, and the oxygen radical disease of prematurity. A.J.D.C., 142, 1341, 1988.