

**Structure and function of a novel
cytochrome c_4 from the purple
photosynthetic bacterium
Thiocapsa roseopersicina.**

Rui Miguel Mamede Branca

Summary of Ph.D. Thesis

Supervisor: Dr. Csaba Bagyinka

Institute of Biophysics, Biological Research Center
of the Hungarian Academy of Sciences

Ph.D. School of Physics
University of Szeged

Szeged

2008

Introduction

In the present work, we discuss the discovery and study of a novel cytochrome *c* from the purple sulfur bacterium *Thiocapsa roseopersicina*. We have characterized the cytochrome through a combination of absorption spectroscopy and circular dichroism (CD) with redox potentiometry and also by using differential scanning calorimetry (DSC). The data gathered place this protein in the c_4 class of cytochromes. We have used Mass Spectrometry to determine its primary structure, which confirmed this cytochrome to be of the c_4 type. It is the first purified and identified cytochrome c_4 from an anaerobic phototrophic bacterium.

Aims of the study

To purify and characterize proteins with redox centers from purple sulfur photosynthetic bacteria in order to shed some light over the electron transport pathways within the metabolism of these organisms.

Particularly, to characterize a novel periplasmic soluble cytochrome *c* from *Thiocapsa roseopersicina*:

- To determine its primary structure, by using the recent advances in Mass Spectrometry.
- To determine the heme content of the cytochrome, and the respective midpoint potential of each redox site.
- To determine its sensitiveness to oxygen, given that the organism is photosynthetically active only under anaerobic conditions, but can survive in aerobic conditions; and to determine its thermal stability, given that temperature is another sensitive parameter to the survival

of the organism. In sum, to elucidate the interrelation of these two parameters in their influence on the cytochrome structure.

- To elucidate the phylogenetics of this protein.

Finally, to use all of the gathered structural information in order to clarify the function of this cytochrome and on a broader sense to gain insight into the electron transport chains of phototrophic bacteria.

Materials and Methods

The protein components from the cells of *Thiocapsa roseopersicina* strain BBS were extracted in a procedure that employed cold acetone. A first round of anion exchange batch liquid chromatography was followed by four rounds of alternating hydrophobic and anion exchange fast protein liquid chromatographies (FPLC). The purity and molecular weight were determined by SDS–PAGE.

The pyridine hemochromogen method was used, in combination with the Bradford method, to determine the heme content and the exact protein concentration.

In preparation for mass spectrometry, four different digest sets were obtained by digestion of the protein with combinations of three different proteolytic enzymes. All measurements were performed on a linear IT-FTICR hybrid instrument. In most measurements, the proteolytic fragments of each digestion set were separated by online gradient reversed-phase micro-Liquid Chromatography prior to NSI, full-scan mass spectra were recorded in the FTICR unit, and MS/MS product ion scans in the IT. Most peptide sequences were determined by interpretation of the MS/MS spectra with the aid of a computer algorithm.

The redox potential of the cytochrome was determined by potentiometric titration using a home-made stirred spectroelectrochemical cell which included a three electrode system. The enforced solution potential between the working and the platinum electrodes was provided by a potentiostat.

CD spectra were measured both in the far-UV range (190–250 nm) and near-UV–visible range (250–700 nm) as a function of the temperature in both anaerobic and aerobic conditions. CD spectra in the range 190–250 nm were used for calculation of secondary structures.

UV-VIS absorbance measurements and DSC measurements were performed to provide additional information in temperature dependence studies.

Results and Discussion

Cytochrome c_4 contains two hemes per molecule, as determined by the pyridine hemochromogen method, and later confirmed by Mass Spectrometry.

After analysis by MS of the peptide mixtures resulting from the proteolytic digestions, we have compiled the most abundant ions into comprehensive lists of peptide masses. Each peptide was then individually sequenced by interpretation of the respective MS/MS data. It was fairly straightforward to determine the sequence of peptides up to 14 amino acids in length (which translates to roughly 1500 Da in peptide mass). Larger peptides posed mounting difficulties. Linking up yielded the complete amino acid sequence of cytochrome c_4 from *Thiocapsa roseopersicina*. To our knowledge, this is currently the largest protein that has been completely sequenced by mass spectrometry alone.

TDGHQAAAPQ VGDPQAGEAK ANGVCLACHG PQGNSLVPLW
 PKLAGQHPEY IVKQLMDFKQ RRANEQMTPM AMPLTDQEV
 DLAAYYATQP KTPGAADPEL ASKGESLYRW GNPETGVPAC
 SGCHGPAGGA GQSLAKFPRL SAQHADYTKQ TLEHFRGALR
 ANDPNGMMRG AAARLSDQEL AAVSQYLQGL SQ

The direct mass spectrometry strategy used in this work can be often faster and more straightforward than the indirect gene sequencing approach.

Cytochrome c_4 proved to be a heat-tolerant protein if maintained under anaerobic conditions. We clarified that oxygen initiates an irreversible unfolding of the protein at high temperatures, most likely through direct binding to the heme's sixth coordination site, which was left vacant after dissociation of the native methionine ligand. It was shown that the methionine-Fe bond (i. e. the 6th axial ligation) is closely linked to the protein moiety (i.e. the protein's secondary structure) and plays a crucial role in the overall folding of the protein.

Cytochrome c_4 proteins are thought to participate in aerobic respiratory pathways, in a position close to the terminal oxidase of the electron transport chain. The discovery of such a cytochrome in an anaerobic photosynthetic organism throws doubt upon this assumption. In *T. roseopersicina*, cytochrome c_4 must participate in photosynthesis instead. More specifically, since the redox titration revealed midpoint potentials of 237 ± 5 mV (1st heme) and 268 ± 6 mV (2nd heme), we suggest to place this cytochrome in the position of electron transport between cytochrome b/c_1 and the tetraheme cytochrome of the reaction centre.

Our phylogenetic survey revealed that cytochrome c_4 is widespread across γ -proteobacteria, and thus it is likely to have this role of periplasmic electron carrier in several species of photosynthetic bacteria.

In purple non-sulfur bacteria, monoheme cytochrome c_2 usually carries out this function, hence it is appealing to speculate that purple sulphur bacteria use instead a diheme cytochrome c_4 in order to link sulphur metabolism with the photophosphorylation cycle.

Summary of novel findings

- We have purified a novel cytochrome from the organism *Thiocapsa roseopersicina*. This was the first purified and identified cytochrome c_4 from a photosynthetic bacterium.
- We have determined the primary structure of cytochrome c_4 from *T. roseopersicina*. This was the largest protein (about 21 kDa in size) whose amino acid sequence has been determined solely by Mass Spectrometry.
- Using recent developments in the field, we have implemented a protein sequencing methodology that allows high-resolution Mass Spectrometry to compete with indirect DNA-based methods.
- We have characterized cytochrome c_4 from *T. roseopersicina* in terms of UV-Vis and CD spectroscopy, correlating spectral profiles to structural features of the protein.
- We have determined the midpoint redox potential of the two heme groups of cytochrome c_4 from *T. roseopersicina*.
- We have discovered that cytochrome c_4 is heat tolerant and indeed shows a very resilient secondary structure (especially the reduced

state) if under anaerobic conditions, which are the normal growth conditions for *T. roseopersicina*. If under aerobic conditions and in oxidized state, the secondary structure and thus the overall fold of the protein is irreversibly affected by heating in a process where oxygen binding to the vacant 6th axial positions of the heme groups plays a decisive role.

- Our survey through the phylogenetics of cytochrome *c*₄ from *T. roseopersicina* has revealed that this type of cytochrome is widespread in γ -proteobacterial organisms, particularly in Oceanospirillales and Chromatiales orders.

- Up until now cytochromes *c*₄ were believed to participate in oxygenic electron transport chains. In contrast, the work here presented indicates that in purple sulphur bacteria (Chromatiales), cytochrome *c*₄ plays a role in anaerobic phototrophic electron transport chains instead.

List of Original Publications

List of publications directly related to the thesis:

1. **Branca RMM**, Bodó G, Bagyinka C, Prokai L. (2007) De novo sequencing of a 21-kDa cytochrome c_4 from *Thiocapsa roseopersicina* by nanoelectrospray ionization ion-trap and Fourier-transform ion-cyclotron resonance mass spectrometry. *J Mass Spectrom* **42**, 1569-1582.
IF = 3.574
2. **Branca RMM**, Bodó G, Várkonyi Z, Debreczeny M, Ősz J, Bagyinka C. (2007) Oxygen and temperature-dependent structural and redox changes in a novel cytochrome c_4 from the purple sulphur photosynthetic bacterium *Thiocapsa roseopersicina*. *Archives Biochem Biophys* **467**, 174-184.
IF = 3.152
3. Tomcová I, **Branca RMM**, Bodó G, Bagyinka C, Smatanová IK. (2006) Cross-crystallization method used for the crystallization and preliminary diffraction analysis of a novel di-haem cytochrome c_4 . *Acta Cryst F* **62**, 820-824.

Other publications:

1. Ősz J, Bodó G, **Branca RMM**, Bagyinka C. (2005) Theoretical calculations on hydrogenase kinetics: explanation of the lag phase and the enzyme concentration dependence of the activity of hydrogenase uptake. *Biophys J* **89**, 1957-1964.
IF = 4.507

Conference proceedings:

1. **Branca RMM**, Bodó G, Bagyinka C, Prokai L. (2007) De Novo sequencing of a 21-kDa Cytochrome c_4 from *Thiocapsa roseopersicina* by Nano-ESI Ion Trap and FT-ICR Mass Spectrometry. *25th Informal Meeting on Mass Spectrometry, Nyíregyháza-Sóstó, Hungary*; p 33
2. **Branca RMM**, Bodó G, Bagyinka C, Prokai L. (2007) De Novo sequencing of a 21-kDa Cytochrome c_4 from *Thiocapsa roseopersicina* by Nano-ESI Ion Trap and FT-ICR Mass Spectrometry. *Regional Biophysics Conference, Balatonfüred, Hungary*; p 74
3. **Branca RMM**, Varkonyi Z, Bodó G, Ősz J, Debreczeny M, Bagyinka C. (2006) Cytochrome c_4 from *Thiocapsa roseopersicina*. *8th International Conference on Membrane Redox Systems, Szeged, Hungary*; p 24

Acknowledgements

I am extremely grateful to my supervisor, Csaba Bagyinka, for his permanent availability, for reinforcing my conviction that physical work is also extremely important (I literally had to cart bricks and lumber towards my PhD), and above all, for treating me as an equal peer in all scientific discussions.

I would like to thank my colleagues in the Biological Research Center, Judit Ősz, who assisted me greatly when I first came to Szeged, Rózsa Verebély, for her kind and ever present assistance, and Gabriella Bodó, for the solidarity through the long days of hard work in the laboratory.

Special thanks to Várkonyi Zsuzsanna and her family, for the limitless hospitality, and for the invaluable help in difficult situations.

Many thanks to László Prokai in Fort Worth, Texas, USA, for teaching me about the enormously powerful method that is Mass Spectrometry.

To my parents, who gave me continuous support from the far edge of Europe, and to my brother Fernando who daringly crossed the continent bridging Portugal to Hungary.

To Erika Bereczki, who helped me through the decisive moments of this laborious and arduous process, summed up in three letters: PhD.

I am grateful to the Institute of Biophysics of the Biological Research Center in Szeged for hosting me through these years of scientific endeavour.

I want to thank the financial support to our laboratory from the Hungarian Science Foundation (grants OTKA T049276 and OTKA T049207), and from AUTOESKORT Ltd.

Finally, I am greatly indebted the Portuguese Science and Technology Foundation for providing me personally with the PhD scholarship (under POCTI, SFRH/BD/13128/2003). May the Foundation continue with their fundamental contribution to the advancement of Science in Portugal.